UNIVERSITY OF MARYLAND EXTENSION

TPM/IPM Weekly Report
for Arborists, Landscape Managers & Nursery Managers

Commercial Horticulture **August 7, 2026**

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Integrated Pest Management for Commercial Horticulture
extension.umd.edu/ipm

If you work for a commercial horticultural business in the area, you can report insect, disease, weed or cultural plant problems **(include location and insect stage)** found in the landscape or nursery to sklick@umd.edu

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Disease Information: David Clement (Extension Specialist) and Ana Cristina Fulladolsa (Plant Pathologist and Director, UMD Diagnostic Lab)
Weed of the Week: Kelly Nichols and Nathan Glenn, (UME Extension Educators) and Dan Buonaiuto, (Assistant Professor), Dept. of Plant Sciences and Architecture
Cultural Information: Ginny Rosenkranz (Extension Educator, Wicomico/Worcester/Somerset Counties)
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No Report on August 14th

Due to vacation schedules, there will not be an IPM report next week on August 14th. We will send out the next report on August 21, 2026.

IPM Diagnostic Session (1 - 3 p.m.)
August 26, 2026
Location: CMREC, Ellicott City, MD

Blister Beetles

By: Holly Greenberg and Suzanne Klick, UME

Blister beetles emerge in late summer/early fall with only one generation per year. The four most common types, among the 250 species present in the U.S., are the striped, ash gray, margined, and black blister beetle. This week, John Austin, McHale/Green Gardens, found the margined blister beetle, which can be identified by the gray margin lining its wings, causing significant damage to anemones. Most are between ½" and ¾" in length, with abdomens extending past their soft wings.

They feed on flowers, including asters, calendulas, zinnias, and dahlias, and trees, including hawthorns and serviceberries. As adults, they are herbivorous and will defoliate the plant while also consuming its pollen and nectar. Eggs are deposited in the soil in late summer. Once the young larvae hatch, they hunt for a patch of grasshopper eggs where they will become immobile grubs until pupation. The larvae are predators that prey on grasshopper eggs, which are pests. The following summer, the grubs pupate and emerge as adults. Blister beetle populations can vary depending on the dryness and number of grasshoppers present the previous summer.

Avoid contacting margined blister beetles without thick gloves.
Photo: Suzanne Klick, UME

The blood of this insect contains the toxin cantharidin, which causes blistering upon contact with human skin and gastrointestinal issues when ingested by both humans and animals. Livestock commonly ingest the chemical through contaminated bales of hay. Only male beetles produce this compound, making mating status the determining factor in whether females contain it.

Blister beetles can show up suddenly in an area and cause damage before moving elsewhere. Control is not always required since they disappear so quickly. Neem can be used as a repellant. Other control options include spinosad, peremthrin, and acephate. A gloved hand or tool can be used to pick individuals off the plant to place them into soapy water.

Wheel Bugs

By: Suzanne Klick

While looking for predators for the Gill Symposium this week, Paula Shrewsbury and Holly Greenberg found an adult wheel bug at our research center. During the lab session, the wheel bug had a spotted lanternfly meal. Wheel bugs are generalist predators with piercing and sucking mouthparts. The wheel bug pierces its prey to inject an enzyme which liquifies their body contents. The wheel bug then uses it sucking mouthpart to feed. There is only generation per year in Maryland. Look for nymphs and adults throughout the summer. Avoid handling wheel bugs because the enzyme they inject into their prey can cause pain in humans.

An adult wheel bug carrying its Japaense beetle meal.
Photo: Suzanne Klick, UME

Next Generation of Box Tree Moth

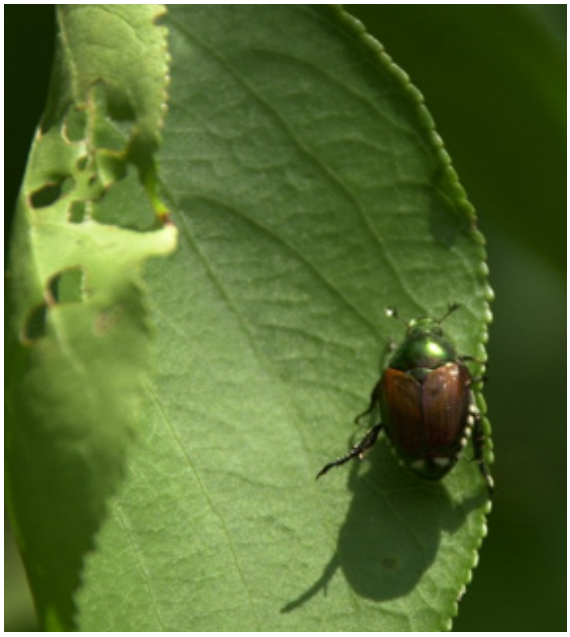
By Laura Nixon and Holly Greenberg

Despite no captures of adult box tree moth (*Cydalima perspectalis*) in our Western Maryland traps over the past 3 weeks, today we found newly hatched caterpillars. These caterpillars were found 0.5 mile down the road from the previous infestation that we were monitoring in Clear Spring, MD. We also found what appeared to be close to hatching eggs. Second instar caterpillars were also found in Williamsport, MD. This is the first time we have seen them at this location, but let us know if you have sighted more in any of the locations mentioned. If you have boxwoods adjacent to infestations, now is the time to monitor for newly laid eggs and hatching larvae. Using *Bacillus thuringiensis* (Bt) at this time is highly effective against the earliest caterpillar life stage. These smaller caterpillars chew at the top leaf layer, as seen in the photo, and leave behind very small webbing and frass piles.

Look for early instar box tree moth caterpillars.
Photos: Holly Greenberg, UME

A later instar box tree moth caterpillar was found at the Williamsport site.
Photo: Holly Greenberg, UME

White Grubs: NOW is the time to monitor for active grubs and feeding damage
By: Paula Shrewsbury, UMD



Japanese beetle adult and skeletonization on the leaf.
Photo: Paula Shrewsbury, UMD

White grubs are the immature stages of scarab beetles (family Scarabaeidae). In this area, the most damaging white grubs are larvae of Japanese and Oriental beetles. We sometimes have large populations of masked chafer grubs, but they tend to be less damaging. White grubs can be a problem in lawn, golf, and nursery turf, and container and B&B nursery stock. The question is "Do we expect white grubs to be bad this year?". Let's start with adult scarab beetle activity. We (UME IPM Team) have not had reports of high levels of Japanese or other scarab beetle densities this year. I have seen a few locations recently (VA Beach VA, Ellicott City MD) where Japanese beetle adults and their damage were relatively abundant. I would describe this as patchy occurrence of scarabs. Let's talk about the weather next. Japanese beetle adults' lay eggs starting around mid-late June through July in the MD area. These eggs need moisture to survive and hatch. Last year, during this time most of the region was under drought conditions which suggests white grub populations should have been low, except maybe in irrigated turf. This would explain the low, patchy occurrence of adult beetles this year. This year, we have been in water deficit since the beginning of the season, and rain has been patchy in its occurrence. Given all of this, I would predict that white grub populations will be low this season. We are dealing with biology so if I am wrong and you are **seeing high populations of adult and/or white grubs, please let me know (which beetles and life stage, and location; pshrewsbury@umd.edu).** I suggest that you play it safe, and that you monitor your turf and nursery stock closely for white grub damage and activity.

White grubs found in the turf feeding on roots.
Photo: P.M. Shrewsbury, UMD

Monitor: If you have observed areas with significant adult scarab beetle activity you should monitor now in those areas to determine if grubs are present. White grubs should be 2nd and early 3rd instars now. Dig soil cores or cut 1 sq. ft. sections of turf in areas where adult scarab beetle activity has been high and/or turf appears drought stressed or discolored. White grubs will be active in the upper few inches of soil in the turf or root zone. Pull on the turf and if it feels loosely attached to the soil you may have white grubs feeding on the turf roots. If you find grubs, identify which species of grubs are in the turf. White grub species can be identified by the rastor (hair) pattern on the underside of the abdomen. A hand lens or some type of magnification will be needed. For help in grub identification go to: http://ohioline.osu.edu/hyg-fact/2000/pdf/2510.pdf

Management: If you have high scarab activity and / or have had historical problems with white grubs, now is an optimal time to treat. The treatment window will only last another few weeks. Once you get past mid-September the effectiveness of products goes way down on later instar grubs. If white grub control is warranted in turfgrass many of the neonicotinoids such as imidacloprid, clothianidin, and thiamethoxam are labeled and give good control. Mach2 is an insect growth regulator that provides good control of grubs. Acelepryn is a relatively newer class of chemicals that has shown excellent control of grubs. Acelepryn has very low toxicity to mammals (no signal word required by EPA in MD) and pollinators (see below).

Ambrosia Gall Midge Spots on Canada Goldenrod

By Laura Nixon

Heather Zindash, The Soulful Gardener, left a gift of goldenrod leaves on my desk this week. These leaves had interesting black and white mottled spots on them from an ambrosia gall midge (*Asteromyia carbonifer*). When adult females insert their eggs into goldenrod leaves, they also inoculate the leaf with a symbiotic fungus. This fungus then forms a gall-like structure for hatching midge larvae to live and develop in. Developing larvae can feed on the fungal material, and it appears that the tougher black structure formed provides physical protection. The midge larvae pupate inside this gall and emerge as fully formed adults. I was able to dissect the *Solidago canadensis* leaves that Heather brought and found orange-colored pupae beneath the fungal layer. If you see these spots forming, I recommend removing and disposing of them; no chemical treatment is necessary.

***Solidago canadensis* leaf with spots from ambrosia gall midge.**
Photo: Laura Nixon, UME

Fungal layer peeled back to reveal midge pupa.
Photo: Laura Nixon, UME

Scout for Magnolia and Tuliptree Scales

By: Laura Nixon

Both magnolia scale (*Neolecanium cornuparvum*) and tuliptree scale (*Toumeyella liriodendri*) have one generation per year, with crawlers becoming active late in the summer. Magnolia scale feeds and reproduces only on magnolia trees, whereas tuliptree scale can be found on magnolias, tulip poplars and lindens. Both species are soft scales, which means they feed on host tree phloem and produce honeydew, which coats plants and propagates the growth of black sooty mold. We expect to see active crawlers hatching from both species in mid-August through to September. Despite the similar biology and overlapping host of these species, they become visually distinctive

Tuliptree scale feeds on both magnolia and tuliptree.
Photo: Suzanne Klick, UME

as the females mature around this time of year if you look closely with your hand lens. Magnolia scale females develop a visually obvious white waxy coating early in the season, which remains as a thinner white layer over an orangey body as the females mature and swell. Whereas tuliptree scale females do not develop the powdery white wax and vary in color from grey to darker orange with a distinct rippled edging (or flange) on the mature females.

Magnolia scale only feeds on magnolia species. Photo: Suzanne Klick, UME

Control: For both species of scale, feeding populations should not affect the vigor of an already healthy tree. Trees undergoing other stressors, such as drought, root damage, or disease, can be affected. For heavily infested trees, the honeydew can be a nuisance and aesthetic issue. Scout now for the visually obvious females to prepare your management plan. Scale bodies can be physically scrubbed away now to minimize the number hatching out next month. When crawlers emerge, these can be treated with insect growth regulators (IGRs) which are less harmful to beneficial insects, such as azadiractin (ex. AzaGuard), pyriproxyfen (ex. Distance) and buprofezin (ex. Talus). Protecting and maintaining natural enemies, including several species of predatory lady beetles, will help keep your scale populations under a damaging level.

Stanton A. Gill Symposium: A Focus on Biocontrol

By: Laura Nixon, Dave Clement, and Suzanne Klick

This week, we hosted the 2nd annual Stanton A. Gill Symposium at the Central Maryland Research and Education Center in partnership with the Maryland Nursery, Landscape and Greenhouse Association (MNLGA). For 2026, we decided to focus on the area of biocontrol, including new technologies and products, the mechanisms of biocontrol, interactions between different control options, and practicalities on how to get started. We were lucky to have such a great line up of guest and UMD speakers. This was definitely reflected by the attendance: sold out, standing room only!

Dr. Brad Fritz, Certis Biologicals, presented on biofungicides and compatibility with tanks mixes on Day 1. Photo: Suzanne Klick, UME

Day 1 opened with Dr. Pam Marrone (Invasive Species Corporation) speaking on the industry of biological control agents, predominantly biorational pesticides. Pam has herself started three biopesticide companies and is responsible for the early development of several biopesticides including Venerate and Serenade. There is a growing market for biopesticide products, which take 4-5 years and approximately $10M to develop for market, compared to conventional chemistries which can take up to 12

years and over $300M to develop. We can expect to see a growing number of efficacious biopesticides for our pest management schedules going forward.

Insect Pest Management with Biocontrols

Dr. Paula Shrewsbury, UMD, covers some insect predators during the lab session on Day 2.
Photo: Suzanne Klick, UME

Dr. Paula Shrewsbury (University of Maryland) taught us all about parasitoids, predominantly wasp and fly species, which lay their eggs in pest species, where their offspring develop at the expense of the pest species. There are many parasitoids present in our natural landscapes, and some that have been harvested for commercial use to release in your crops, including the *Aphidius colemani* wasp. *Aphidius colemani* lays its eggs in aphids, where the larvae develop whilst the aphids remain alive and swell into "aphid mummies" before the adult wasp emerges. To follow up on this lecture, Paula (along with Drs Mike Raupp (The Bug Guy) and Laura Nixon (UME)) hosted a workshop on day 2 focused on mother nature's hit squad. In this workshop, attendees used microscopes to see the aphid parasitoid wasp laying her eggs in live aphids! We also got to see several predators which directly hunt and consume pests, including lady beetles and lacewing larvae.

Dr. Carlos Bográn (Bioworks) gave us a rundown on how biorational insecticides function and interact with classical biocontrols (i.e. predatory arthropods). There are several online tools, many provided by biopesticide vendors, which allow you to check for compatibility between your selected biopesticides and any augmentative predators you have released. There is also an EPA approved scale for the effect of pesticides on other beneficial insects such as pollinators.

To wrap up day 1, Heather Zindash (The Soulful Gardener and MNLGA Education Committee) brought together much of the presented information from the day and taught us about how to physically get started with an effective biocontrol program in your greenhouse or nursery. Step 1: Identify your pest as close to species level as possible! This will guide your selection of biopesticide or augmentative biocontrol releases.
As part of Day 2, Nan McKay and Kristen Fowler (Koppert) presented on some best practices for implementing an effective biological control program. One of the things that they pointed out is the importance of knowing how different beneficial mites interact. For example, if you release *Neoseiulus (Amblyseius) cucumeris* and choose to release *Amblyseius swirskii* as it warms up, you will need to kill the *N. cucumeris*. *A. swirskii* will feed on *Neoseiulus* instead of crop pests that you are trying to control.

Disease Pathogens for Weed Management and Proper Management and Diagnosis of Unwanted Pathogens

In the morning of day 1 Dr. Dan Buonaiuto (UMD), covered the basics of fungal pathogens for weed management and how they could be used for difficult weeds. Especially, where large land areas such as range lands need management, or when herbicide resistance is an issue. The choice of autoecious rust fungi, (single

host), when available gives the highest specificity for weed management since rusts are obligate parasites and have low risk of mutating to non-target plants. Dr Dave Clement (UME) gave an example of Canada thistle management with the rust pathogen *Puccinia punctiformis* in Maryland. We had mixed results with weed control efficacy and explained how we could explore new inoculation techniques in the future.

In the afternoon on day 1 Dr. Brad Fritz (Certis) presented the use cases of bio fungicides and microbial elicitors and how to use them. He stressed early application before disease becomes overwhelming, modes of action, and application intervals as well as compatibility with tank mixes. Some products, especially Bacillus formulations, have multiple modes of action. Systemic acquired resistance and induced resistance describe different plant responses to pathogen attack, and the use of plant defense elicitors results in reduced disease. He also mentioned the importance of using different FRAC groups, just as with conventional fungicides to achieve better control efficacy.

On day 2, Drs. Ana Christina Fulladolsa, Karen Rane, and Dave Clement (UMD) lead a hands-on lab diagnosing plant diseases emphasizing immune test kits for plant viruses that are common in the nursery industry. We emphasized the importance of taking green living tissue and younger plant leaf samples from several plants and separate leaves within the same plant since virus distribution is not uniform within diseased tissue. We covered how critical it was to see a positive test kit line as well as the location of the virus positive line for diagnosis.

We would like to finish by saying a huge thank you to all the support for this event. We appreciate the ongoing support of our Dean Wendy Powers, our partner MNLGA, and our amazing stakeholders (that's you!).

Dr. Ana Cristina Fulladolsa presents on how to use diagnostic test kits.
Photo: David Clement, UME

Spider Mite Activity Continues

With the hot weather, we continue to see spider mite activity. Elaine Menegon, Good's Tree and Lawn Care, found active spider mites on winged euonymus in Lititz, PA this week. Look for yellow stippling damage on the topside of leaves and mites, eggs, and cast skins on the underside of the foliage. There are numerous commercially available miticides but be careful to read the label before applying; not all miticides kill all types of mites. Good options for hitting both spider and eriophyid mites include spiromesifen (mite growth regulator), or acequinocyl (EPA reduced risk product)

This euonymus has heavy stippling damage from spider mite feeding.
Photo: Elaine Menegon, Good's Tree and Lawn Care

Tree Damage Mystery

From Liam Ulasevich, Casey Trees: "I wanted to share this situation even though it's not a pest or disease. A DC resident reached out to Casey Trees asking what had gotten to the base of their tree (see pictures). Our staff primarily guessed beavers, rot, or vandalism. However, neither they nor the resident were completely confident in these answers. Our Field Manager, Nick Smalley, had a hunch that the resident should go back to the tree and look up. She did. Turns out the "tree" was not a tree at all but actually a large branch that fell and spiked the ground.

This scenario reminded my team that when we're interpreting nature, it is helpful to question even the most basic assumption we have about our observations."

A different perspective can help identify plant 'problems'.
Photos: Provided by Casey Trees

Beneficial of the Week

By: Stacy Small-Lorenz, UME

Meet the Smoky-winged Beetle Bandit Wasp (*Cerceris fumipennis*)

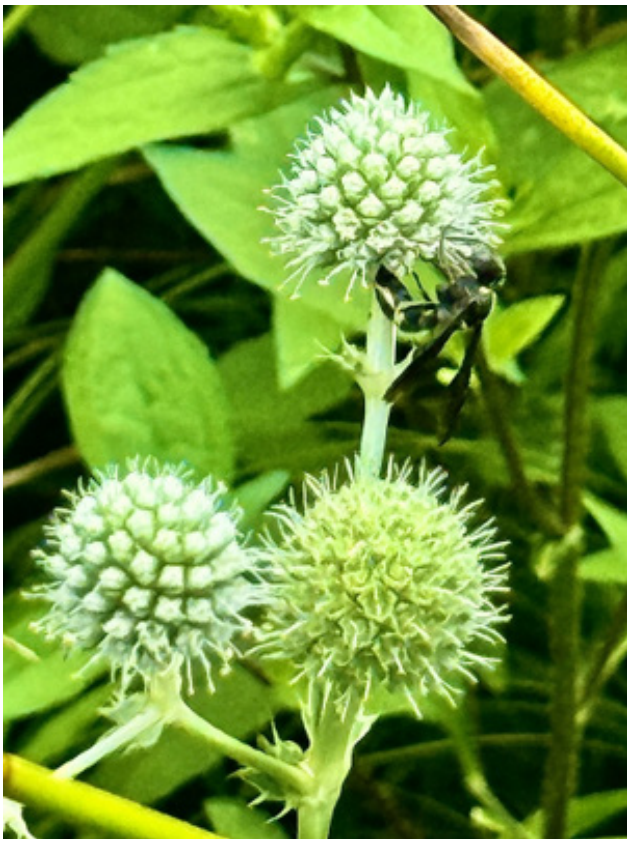
I was transplanting a few blue mistflower (*Conoclinium coelestinum*) plants into my drainage swale pollinator garden last weekend when I noticed this new insect visitor to my blooming rattlesnake masters (*Eringium yuccifolium*; below).

This gentle beauty has a name that could have been penned by a cowboy poet – smoky-winged beetle bandit wasp (*Cerceris fumipennis*; Family Crabronidae) – and it plays an important ecological role. It parasitizes adult metallic wood-boring beetles of the family Buprestidae, including oak-boring beetles and the emerald ash borer (*Agrilus planipennis*), an invasive beetle that is devastating North American ash trees, especially Green and White Ashes (*Fraxinus pennsylvanica* and *americana*).

The smoky-winged beetle bandit wasp is native to Eastern North America and is the only North American wasp known to parasitize *adults* of Buprestid wood-boring tree pests, using the beetle bodies as hosts for their eggs and larvae. Scientists have even used this species for biosurveillance to monitor range expansion of the emerald ash borer.

A combination of traits can help identify this native parasitoid wasp. Look for its black head, thorax, and abdomen and smoky-brown wings. Females may have a blue reflection to their wings. A distinctive cream or white apical band on the second abdominal segment is another key characteristic. Females have three cream-colored spots on the face, while males have two yellow triangles between their eyes.

Smoky-winged beetle bandit wasp adults are active mid-June into September in the northern part of their range. They are solitary ground-nesters that generally produce two generations annually. According to Heather Holm (2021), they usually burrow into compacted sandy or gravelly soil along rivers, in paths, gravel parking lots, or even baseball diamonds, frequently near forests where they can hunt for their beetle victims in the tree canopy, then bring them back to the nest. A female captures and injects a beetle with venom, then carries it in her mandibles to stash with several other paralyzed beetles in an underground nest chamber.



A smoky-winged beetle bandit wasp nectars on a rattlesnake master flower in the author's pollinator garden.
Photo: Stacy Small-Lorenz, UME

She'll lay one egg on the top beetle that hatches after several days. After hatching, her larvae will remain in their underground chambers over winter, feasting on beetles from their own personal stockpiles, before emerging as adults the following summer. She provides a larger supply of beetles for her female offspring than for the males. Biologists can observe the beetles that female wasps bring back to their ground burrows to document the presence of these tree pests.

These beneficial wasps are also "accidental" pollinators that pick up pollen on their bodies and transport it to other flowers while nectaring. They are harmless to humans and defenders of our forests, so two things we can do for them in return are protecting their nest burrows when we find them and providing some of their preferred nectar plants in a sunny spot near forest, like rattlesnake master, swamp or butterfly milkweed (*Ascelpias incarnata or tuberosa)*, Virginia mountain mint (*Pycanthemum viginianum*), daisy fleabane

A female smoky-winged beetle bandit wasp brings a pair of mating emerald ash borer beetles back to her nest. The blue paper shown here will make it difficult for her to bring her victims into the nest, allowing biologists to collect the beetle bodies for pest monitoring.
Photo: Philip Careless, Wikimedia Commons [CC BY-SA 3.0]

(*Erigeron strigosus*) and Canada goldenrod (*Solidago canadensis*). These are relatively simple pest management practices for forest conservation that will also benefit many other pollinators.

While one species of wasp alone may not entirely solve the widespread dilemma of invasive wood-boring tree pests, there are other introduced and native parasitoid wasp species and woodpeckers that target earlier stages of the emerald ash borer life cycle (egg and larval) (Shrewsbury 2026). Therefore, working in combination with other species, they could make a real difference for the next forest generation and therefore deserve our conservation attention.

References:

Holm, H. 2021. Wasps: Their Biology, Diversity, and Role as Beneficial Insects and Pollinators of Native Plants. Pollination Press LLC. Minnesota, USA. 420 pp.

Shrewsbury, P. 2026. The Exciting Life of Parasitoids and Their Role as Biological Control Agents. Presented at the Stanton A. Gill Symposium: A Focus on Biological Control. August 5, 2026. University of Maryland Extension, Clarksville, MD.

Weed of the Week

By: Dr. Dan Buonaiuto

This week's weed of the week is mugwort, *Artemisia vulgaris,* which can be a particularly problematic weed in landscape and nursery settings. This plant was introduced to North America in the 1600's for its medicinal properties.

Figure 1: Mugwort leaves.
Photo: Ohio State Weed Lab, The Ohio State University, Bugwood.org

As part of a larger research project, I have been surveying natural resource managers across the northeast for the past two years about the different invasive plants they manage, and they constantly report that mugwort is one of the most difficult invaders to manage in natural areas. Let's take a closer look at the ecology and management of this species.

Mugwort Identification:

- **Habit:** Mugwort is an herbaceous perennial that spreads though underground rhizomes, forming thick patches, which can range in height from 2-5 feet depending on the growing conditions.
- **Leaves:** Alternately arranged, three to four inches in length and one to three inches in width. Leaves are deeply lobed, and green on the top with dense grey to white hairs on the underside (Figure 1). Leaves are also aromatic.
- **Stems:** Stems are purple to brown, hairy, and become woody with age.
- **Flowers:** The wind-pollinated flowers are inconspicuous and lack petals.

Mugwort Ecology:

- Mugwort grows aggressively in partial to full sun and dry soil, but does not tolerate wet soils (Figure 2).
- This plant spreads aggressively though underground rhizomes, but also reproduces through seed, which are dispersed in the fall.

Mugwort Control:
This species is notoriously difficult to control and may require a combination of several approaches listed below, and take multiple seasons of work.

Mechanical control:

- You can handpull very young plants in the early spring before the rhizomes have formed. Do not try to handpull or till larger patches, as broken roots and rhizomes will generate new plants.

- Smothering mugwort patches with landscape fabric, thick layers of plastic or mulch can also help control patches in open areas.

- Mowing in the late summer can be an effective way to prevent seed production thus reducing the spread potential of existing patches but will not reduce current infestations.

Chemical control:

- The dense hairs on the leaves of mugwort can make it difficult for herbicide to penetrate the leaf surface, and herbicides may be more effective with a surfactant.

- Foliar spray with a systemic herbicide like glyphosate or triclopyr applied in the late summer or early fall will likely reduce the mugwort coverage the following year, but not eliminate patches.

Figure 2: A mugwort patch on the landscape.
Photo: Robert Vidéki, Doronicum Kft., Bugwood.org

Plant of the Week

By: Ginny Rosenkranz

Rosa Jacruwhi 'Wild Spice' is a lovely compact hybrid rugosa rose that only grows 3-4 feet tall and 2-3 feet wide. Each pure white semi double flower spreads 4 inches wide with 5-8 fragrant petals that surround the yellow centers. The flowers have the delightful habit of blooming from May throughout the summer and into autumn's first frost. Butterflies and other pollinators flock to the open flowers and provide extra color and motion in the garden. Rugosa means wrinkled; the green leaves with their strong venation can look wrinkled. After the flowers have finished blooming, the shiny red orange rose hips develop and provide food for many birds and small mammals. The plants are best grown in full sun to partial afternoon sun,


***Rosa* Jacruwhi 'Wild Spice' flower**
Photo: Ginny Rosenkranz, UME

and thrive in medium moist, well drained garden loam. It is always best to water the plants in the mornings and keep the soil lightly moist but not waterlogged. A light summer mulch helps keep the soil moist and cool and reduces the number of summer weeds. The more sun the plants get, the more disease resistant and the more flowers are produced. Plants are cold tolerant from USDA zones 5-8 and can be planted in cottage gardens, as cut flowers and in a hedge. The rose hips are not only beautiful but also edible and safe for both people and pets. Insect pests include aphids, Japanese beetles and thrips. 'Wild Spice' has excellent disease resistance to black spot and mildew.

***Rosa* Jacruwhi 'Wild Spice' plant (above) and rose hips (right)**
Photos: Ginny Rosenkranz, UME

Pest Predictive Calendar "Predictions"

By: Nancy Harding and Paula Shrewsbury

In the Maryland area, the accumulated growing degree days (**DD**) this week range from about **2149 DD** (Clarksville) to **2976 DD** (Nat'l Arboretum/Reagan Nat'l). The Pest Predictive Calendar tells us when susceptible stages of pest insects are active based on their DD. Therefore, this week you should be monitoring for the following pests. The estimated start degree days of the targeted life stage are in parentheses.

Maskell scale – egg hatch / crawler (2nd gen) **(2035 DD)**
Euonymus scale – egg hatch / crawler (2nd gen) **(2235 DD)**
Mimosa webworm – larva, early instar (2nd gen) **(2260 DD)**
Japanese maple scale – egg hatch / crawler (2nd gen) **(2508 DD)**
Fern scale – egg hatch / crawler (2nd gen) **(2813 DD)**
White prunicola scale – egg hatch / crawler (3rd gen) **(3238 DD)**
Spotted lanternfly – egg laying **(Sept.)**

See the Pest Predictive Calendar for more information on DD and plant phenological indicators (PPI) to help you better monitor and manage these pests.

Degree Days (as of August 5, 2026)

Annapolis Naval Academy (KNAK)	2633
Baltimore, MD (KBWI)	2625
Belcamp (FS836)	2526
Clarksville (001MD)	2149
College Park (KCGS)	2713
Dulles Airport (KIAD)	2647
Ft. Belvoir, VA (KDA)	2752
Frederick (KFDK)	2395
Gaithersburg (KGAI)	2534
Greater Cumberland Reg (KCBE)	2352
Martinsburg, WV (KMRB)	2435
Millersville (MD026)	2626
Natl Arboretum/Reagan Natl (KDCA)	2976
Perry Hall (C0608)	2453
Salisbury/Ocean City (KSBY)	2607
St. Mary's City (Patuxent NRB KNHK)	2781
Westminster (KDMW)	2749

Important Note: We are using the Online Phenology and Degree-Day Models site. Use the following information to calculate GDD for your site: Select your location from the map Model Category: All models Select Degree-day calculatorThresholds in: Fahrenheit °F Lower: 50 Upper: 95 Calculation type: simple average/growing dds Start: Jan 1

Conferences

IPM Diagnostic Session (1 - 3 p.m.)
August 26, 2026
Location: CMREC, Ellicott City, MD

FALCAN Truck & Trailer Safety Seminar
October 14, 2026
Location: Urbana Fire Hall, Urbana, MD
Registration opens in July.

2027 Advanced Landscape IPM PHC Short Course
This is a recertification short course for arborists, landscapers, IPM consultants, horticulturalists, professional gardeners, and others responsible for urban plant management. The course lectures will be held over four days at the University of Maryland, College Park, MD. In addition, there will be a hands-on lab following lecture (available to a limited number of course attendees). Coordinators: Drs. Paula Shrewsbury and Mike Raupp, Dept. of Entomology, University of Maryland
Lecture dates: Monday, January 4 - Thursday, January 7, 2027 from 8:00 am – 3:00 pm
Lab dates: Monday, January 4 - Thursday, January 7, 2027 from 3:30 pm – 5:30 pm (space limited)
Course information: https://landscapeipmphc.weebly.com/
Registration: https://www.eventbrite.com/e/2027-advanced-landscape-ipm-phc-short-course-tickets-1992372702386?aff=oddtdtcreator

Questions contact: Amy Yaich, 301-405-3911, umdentomology@umd.edu

Commercial Ornamental IPM Information
http://extension.umd.edu/ipm

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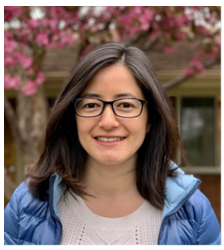

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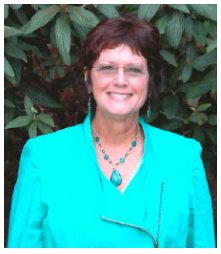

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