UNIVERSITY OF MARYLAND EXTENSION

TPM/IPM Weekly Report

for Arborists, Landscape Managers & Nursery Managers

Commercial Horticulture **July 17, 2026**

In This Issue...

Integrated Pest Management for Commercial Horticulture
extension.umd.edu/ipm

If you work for a commercial horticultural business in the area, you can report insect, disease, weed or cultural plant problems **(include location and insect stage)** found in the landscape or nursery to sklick@umd.edu

Coordinator Weekly IPM Report:
Laura Nixon, IPM Specialist in Entomology, University of Maryland Extension, Lnixon1@umd.edu, and Paula Shrewsbury, Professor and Extension Specialist in Ornamental and Turf IPM, Department of Entomology, pshrewsbury@umd.edu

Regular Contributors:
Pest and Beneficial Insect Information: Laura Nixon and Paula Shrewsbury (Extension Specialists) and Nancy Harding, Faculty Research Assistant
Disease Information: David Clement (Extension Specialist) and Ana Cristina Fulladolsa (Plant Pathologist and Director, UMD Diagnostic Lab)
Weed of the Week: Kelly Nichols and Nathan Glenn, (UME Extension Educators) and Dan Buonaiuto, (Assistant Professor), Dept. of Plant Sciences and Architecture
Cultural Information: Ginny Rosenkranz (Extension Educator, Wicomico/Worcester/Somerset Counties)
Fertility Management: Andrew Ristvey (Extension Specialist, Wye Research & Education Center)
Design, Layout and Editing: Suzanne Klick (Technician, CMREC)

Stanton A. Gill Symposium: Focus on Biological Control
August 4 and 5, 2026
Location: CMREC, Ellicott City

MDA Pesticide Recertification Approval:
Categories 3A, 3B, 3C, 10, CORE, PVT

Virginia Pesticide Recertification Approval:
3A (23), 10 (10), and 60 (60)

Schedule submitted to DC for approval

Spotted Lanternfly (SLF) Adults

We are starting to receive reports of spotted lanternfly adults active in the area. Late instar nymphs also continue to be reported. Both of these stages of SLF produce heavy amounts of honeydew. Look for sooty mold growing on the honeydew.

July 22, 2026 IPM Diagnostic Session
Time: 12:30 - 3:00
Location: CMREC, Ellicott City

The link to register for this program is on our Conferences' web page

This session is a great opportunity to bring plant samples with problems you need identified. Entomology, plant pathology, and horticulture extension specialists will be available to look at samples under the microscope and answer questions. There will be short presentations on tarnished plant bugs and another timely insect pest.

Box Tree Moths are Flying Again

By: Laura Nixon

In Clear Spring, MD this week, we found new empty pupal casings, which means box tree moth (*Cydalima perspectalis*) have developed into a new generation of adults. Adults can fly and disperse to lay their eggs; if you live in Washington County, keep an eye out over the next few weeks for new infestations on your boxwoods. You can also monitor for adult moths with traps (see the May 8, 2026 IPM Report for trap information). At this point, any caterpillars on the plants are quite large and preparing to pupate. Sprays are less effective on these larger caterpillars, so I would recommend picking off caterpillars you see and pruning out branches with signs of activity. If you have a heavy infestation, you can follow this up with an insecticide application (see the June 12, 2026 IPM Report for recommendations).

Empty pupal casings are brown. The black head capsule is left over from when the box tree moth caterpillar pupated, suggesting this is a fresh pupal case.
Photo: Laura Nixon, UME

Rusts and Slime Molds

By: David L. Clement

Cedar-quince rust, *Gymnosporangium clavipes*, will infect the fruit of Callery pear. This fungal pathogen also infects other woody plants in the rose family such as hawthorn, quince, serviceberry, pear, crabapple, apple, and mountain ash. The alternate hosts include several species of juniper. If your clients park cars under infected trees, the red/orange spores can accumulate on the cars.

Another rust that attacks Callery pear later in the fall is Pear Trellis rust caused by *Gymnosporangium sabinae*. It attacks the leaves and causes brown, blister-like swellings on the lower leaf surfaces. This infection can cause defoliation. It also has several juniper species as the alternate hosts.

Pear trellis rust causes brown, blister-like swellings on the lower leaf surface.
Photo: David Clement, UME

Slime molds will be visible after summer rains on mulch and turf at this time of year. These are non-pathogenic organisms that feed on microbes and organic matter in the soil/mulch substrate and then coalesce into fruiting structures on the surface to spread their spores. The slime molds on turf tend to be gray, or black and commonly belong to either the genera *Mucilaga*, or *Physarum*. The slime mold common on mulch is usually *Fuligo septica.*

Slime mold in a mulched area: the crust made up of calcium carbonate was broken open to show black spores within.
Photo: Suzanne Klick, UME

Grass blades covered with slime mold have a darker appearance due to the black spores within the crust.
Photo: Suzanne Klick, UME

Still Seeing Small Bagworms

By: Laura Nixon

Marie Rojas, IPM Scout, sent us photos of small bagworms this week. These particular bagworms borrowed some of Marie's flagging tape to decorate their bags!

If you are still seeing small bagworms, they can be picked off. If you are seeing high populations of bagworms, the smaller caterpillars are much more susceptible to treatments. See the May 22, 2026 IPM Report for monitoring and treatment recommendations.

Bagworms use debris from the tree to form their bags.
Photo: Marie Rojas, IPM Scout

Obscure Scale

By: Suzanne Klick

Marie Rojas, IPM Scout, is finding settled nymphs of obscure scale this week. Check to see if crawlers are still active on trees. Monitor trees closely next year in early June for the start of the crawler period to treat at the most susceptible stage. If densities of scale and or damage are high, consider dormant oil applications.

Obscure scale is in the settled crawler stage this week.
Photo: Marie Rojas, IPM Scout

Oak Flake Gall

By: Laura Nixon

These fuzzy white galls are formed on the underside of oak leaves by the cynipid oak flake gall wasp (*Neuroterus quercusverrucarum* syn. *N. floccosus*). This wasp has one generation per year, with each gall containing a single wasp larvae which develops through the summer. The galls are white early in the season and darken over time to brown before the leaves drop. Galls are visible on the upper leaf surface as lumpy growths. These galls don't harm the tree and don't require treatment. If there is an aesthetic concern, leaves can be pruned out.

Oak flake galls on the underside of an oak leaf.
Photo: Marie Rojas, IPM Scout

View of the topside of an oak leaf with oak flake galls.
Photo: Marie Rojas, IPM Scout

Giant Leopard Moth

By: Suzanne Klick

Marlee Muchinsky, Good's Tree and Lawn Care, found a giant leopard moth (*Hypercompe scribonia*) in Camp Hill Park, PA on July 17. In late summer into fall, you might come across the large, black and red caterpillar of this moth. It curls up when disturbed showing its bit of red coloring. It feeds on a variety of plant material, but is not found in large numbers to be a plant pest.

There is another similar moth called the leopard moth, *Zeuzera pyrina*. It has solid spots, not the open rings like on the giant leopard moth. The leopard moth caterpillar is found boring into trees and can be a problem. It is yellow or orange with a black head and black dots along its body.

The giant leopard moth overwinters in the larval stage.
Photo: Marlee Muchinsky, Good's Tree and Lawn Care

Mimosa Webworms

By: Suzanne Klick

Elaine Menegon, Good's Tree and Lawn Care, found active mimosa webworms on honey locust in Lancaster, PA on July 17. There are two generations in Maryland. Damage from the second generation occurs in August into September and is usually more severe. Caterpillars web the leaves together and feed within this nest for protection. Check for the stage of caterpillars. Bt can be used when larvae are small. Spinosad is another control option. Be sure to get good coverage when making applications. If you miss treating trees this year, be sure to monitor honey locust next year in early June at the start of the first generation.

Mimosa webworms can cause significant damage to honey locust trees.
Photo: Elaine Menegon, Good's Tree and Lawn Care

Gloomy Scale Update

By: Paula Shrewsbury, UMD

In the last week Marie Rojas (IPM Scout) in Gaithersburg and Nancy Harding (UMD) in College Park, MD monitored red maples for gloomy scale (*Melanaspis tenebricosa*, Diaspididae). They found that crawler activity is over, and that most of the populations were settled first instars and some later instars. At this time treatments with insect growth regulators would not give good control. For those with gloomy scale infestations, consider dormant oil applications at the appropriate time. Be sure to monitor for crawler activity in the spring.

Gloomy scale infestation on 7/14/2026 on a red maple in Gaithersburg, MD. At this time it appears crawler activity is over and there are most capped first instars and likely some later instars present.
Photo: Marie Rojas, IPM Scout

Warm Weather Mites are Ramping Up

By: Laura Nixon

The hot weather over the past few weeks is resulting in an uptick in spider mite damage. When the weather is hot and dry, spider mites are able to reproduce every 5-10 days, causing populations to explode. Look out for fine webbing and leaf stippling as signs of spider mites; heavy feeding damage causes leaves to curl, brown, and drop. Marie Rojas, IPM Scout, sent us a photo of spider mite damage on an oak leaf from a nursery this week. If you flip over leaves with suspected mite damage, you can see the populations on the underside, as we did whilst scouting a different nursery. Using a hand lens, we were able to identify that there were two-spotted spider mites on the underside of lion's ear (*Leonotis leonurus*) leaves. Scout for mites on both woody and herbaceous plants and treat whilst populations are low when possible. There are numerous commercially available miticides but be careful to read the label before applying; not all miticides kill all types of mites. Good options for hitting both spider and eriophyid mites include spiromesifen (mite growth regulator), or acequinocyl (EPA reduced risk product).

Spider mite damage on oak.
Photo: Marie Rojas, IPM Scout

Two-spotted spider mite adults and eggs on salvia.
Photo: Suzanne Klick

Yellow stippling damage occurring on the lower foliage of *Leonotis leonurus* foliage.
Photo: Suzanne Klick

Redhumped and Yellownecked Caterpillars are Active

By: Paula Shrewsbury, UMD

Yellownecked caterpillars (*Oedemasia concinna*) on oak and redhumped caterpillars (*Oedemasia concinna*) on *Nyssa sylvatica* were reported by Marie Rojas (IPM Consultant) this past week. Both caterpillars belong to the prominent moth family, Notodontidae.

Early instar yellownecked caterpillars clustered on an oak leaf.
Photo: Marie Rojas, IPM Scout

Yellownecked caterpillars are mid-late season (~July – October) defoliators and have one generation per year and overwinter as pupae. They have a wide host range but prefer shade and fruit trees, birch, elm, oak, maple, walnut, mountain ash, and honeylocust. Yellownecked caterpillars feed gregariously (in groups) and are often observed feeding in clusters near the terminal ends of trees. Feeding by young caterpillars skeletonize leaves and larger caterpillars' defoliate branches. When they defoliate one branch, they will move to another to feed.

Redhumped caterpillars is also a mid-late season defoliator that has a similar life cycle and causes similar damage to trees as the yellownecked caterpillar. They overwinter as last instar larvae in coccons (prepupal larvae) in the ground. Handling redhumped caterpillars may cause skin irritation, so be wary.

Management:
Most often control of these caterpillars is not necessary. There are numerous parasitic wasps and flies (ex. tachinid flies), and predators that attack these caterpillars. Laura Nixon found redhumped caterpillar that had been parasitized by a wasp in Boonsboro, MD this week. See the image of the caterpillar mummies. Look for beneficial insect activity to help determine if any or which treatments are necessary. The best control is to hand-remove or prune out branches with clusters of caterpillars. If other options are warranted, control is best and there are more options when caterpillars are in the early-instar stages. *Bacillus thuringiensis* (B.t.), spinosad, horticultural oil and soap (read cautions related to heat and humidity), and the insect growth regulator, tebufenozide (Confirm) work well. Chlorantraniliprone (Acelepryn) or cyantraniliprole (Mainspring) should also work well.

Late instar yellownecked caterpillars. Note the diagnostic yellow colored segment behind the black head.
Photo: Joe Boggs, OSU Extension

Late instar redhumped caterpillars. Note the red colored segment that is diagnostic for this caterpillar.
Photo: Marie Rojas, IPM Scout

Redhumped caterpillar mummies that have been parasitized by a parasitoid wasp.
Photo: Laura Nixon, UME

Tachinid fly parasitoid adult and yellownecked caterpillar with white eggs from the tachinid on its head.
Photo: Suzanne Klick

Powdery Mildew

We will continue to see powdery mildew infection during the humid weather this summer. Marie Rojas, IPM Scout, is finding this disease on both flowering dogwood and crape myrtle everywhere she scouts.

See David Clement's article on powdery mildew management in the July 10, 2026 IPM Report.

Powdery mildew infection on flowering dogwood.
Photo: Marie Rojas, IPM Scout

Ambush Bug: Insects beware

By: Paula Shrewsbury, UMD

This week Marie Rojas, IPM Consultant, found a jagged ambush bug (*Phymata* sp.) in a flower head of mountain mint. The ambush bug is a highly camouflaged sit and wait predator and is usually found hanging out in flowers waiting for prey to come close. See the Beneficial of the Week (Sept. 6, 2024) on ambush bugs for more information on this beautiful and interesting predator.

Ambush bug predator hiding in a mountain mint flower head.
Photo: Marie Rojas, IPM Scout

An ambush bug, a sit and wait predator, with a honey bee nearby that may be its next meal.
Photo: Marie Rojas, IPM Scout

Ants Farming Aphids and Scale

By: Holly Greenberg, UME

This week, Marie Rojas (IPM Scout) observed ants farming aphids on *Nyssa sylvatica.* The ants and aphids are engaged in a mutualistic relationship, meaning both parties benefit. The aphids use their sucking mouthparts to access and consume phloem sap from plants, leading to their excretion of a honeydew substance. The ants rely on this substance as a primary food source and, in turn, protect the aphids from predators and carry them to fresh plant material. The ants use their antennae to stroke the aphid's back and stimulate their release of honeydew. At times, the relationship is no longer equally beneficial as the ants remove the wings and decrease the walking ability of aphids to increase honeydew production near the ants' nest. The gradient of this symbiotic relationship can vary, as many ants only eat the honeydew, while others alter the aphids' bodies to cater to the ants' needs.

Ants tending to aphids on *Nyssa sylvatica*.
Photo: Marie Rojas, IPM Scout

Similarly, Sheena O'Donnell (UME) witnessed ants farming and protecting Crape Myrtle Bark Scale this week. The ants constructed shelters made of dirt and small tree pieces for the scale, reducing predation and exposure to the elements. The ants mobilize the scale by carrying the insect on their back to fresh feeding sites, ensuring continuous honeydew production. The relationship between the scale and ants can lead to a rapid health decline for the plant. As the ants feed on the honeydew from the scale, the sooty mold population increases, reducing the plant's ability to photosynthesize, ultimately leading to its death.

Ant populations can be reduced by moving nests, cutting off branches that act as bridges between the ground and the plant, or spraying dish soap on the plant to remove both the ants and honeydew. By limiting any interactions, the number of natural enemies will increase, leading to a decrease in the scale population.

Ant-built structure protecting scale on crape myrtle tree.
Photo: Sheena O'Donnell, UME

Another closer view of the structure surrounding crapemyrtle bark scale.
Photo: Sheena O'Donnell, UME

Botryosphaeria Dieback on Stressed Woody Ornamentals

By: Ana C. Fulladolsa, Sheena O'Donnell, and David L. Clement

In the last few weeks, we have received several samples of woody ornamentals showing symptoms of branch dieback and cankers in the Plant Diagnostic Laboratory. In most cases, we found woody tissue infected with the fungal pathogen *Botryosphaeria* sp. The pathogen can cause severe symptoms on plants stressed by drought and freeze damage, prevalent factors affecting the landscape this year.

Botryosphaeria canker on branches of Eastern redbud.
Photo: Mary Ann Hansen, Virginia Polytechnic Institute and State University, Bugwood.org

The pathogen is spread by wind, rain, and contaminated tools, entering the plant through natural openings such as lenticels and growth cracks in the bark. Infected plants may have wilting foliage that remains attached on scattered dying branches. Cankers under the bark turns the killed cambium black, or brown. Dead bark can become roughened and cracked as the fungal fruiting bodies emerge. Cankers can enlarge during the growing season and girdle entire branches, or trunks.

Prevent infection by minimizing drought stress and by choosing appropriate planting sites. Remove diseased branches at least 6-8 inches below visible cankers. Sanitize tools between uses, refraining from operating heavy equipment close to trunks such as lawnmowers, and removing diseased branches.

***Botryosphaeria* sp. fruiting bodies (small, black pycnidia) on rhododendron branch with dieback symptoms.**
Photo: Penn State Department of Plant Pathology & Environmental Microbiology Archives, Penn State University, Bugwood.org

Beneficial of the Week

By: Paula Shrewsbury

Parasitoids and their diverse biologies and lifestyles

Parasitoids are a diverse group of insects, mainly in the orders Hymenoptera (wasps) and Diptera (flies), that provide the ecosystem service of biological control of pest insects in natural and managed environments. A few species of beetles, twisted wing insects, moths, and other insects also have been identified as parasitoids. An insect **parasitoid** is an organism that spends its larval stage within or attached to the outside of the body of another insect, known as its **host**, where it develops and feeds, and ultimately results in death of the host. Parasitoids mainly leave their host only as adults when they feed, mate and find new hosts to lay (oviposit) their eggs. Parasitoids differ from parasites, such as ticks or bed bugs, in that parasites feed on their host but usually do not kill it.

***Trissolcus* wasp inserts her eggs into the eggs of stink bugs.**
Photo: Jack Kelly Clark, UC IPM

There are around 68,000 species of named parasitoids worldwide with an estimate of over 700,000 species still undescribed. Parasitoid wasps range in size from 0.008" (ex. the fairyfly family Mymaridae; *Anagrus takeyanus* parasitoid of lace bug eggs) to 5" (ex. the family Ichneumonidae; *Megarhyssa* wasp species that have long ovipositors and attack wood boring larvae) long. Most **wasp** parasitoids use their ovipositor to insert eggs into or lay eggs on their hosts. **Fly** parasitoids do not have ovipositors, and they usually glue their eggs onto the body of their host which hatches and bore into the host, or on leaves that the host consumes and the egg hatches inside the host. Parasitoids are known to attack all life stages (egg, immature, pupa, or adult) of arthropods, but a given parasitoid species usually attacks a specific life stage of a host (ex. *Trissolcus japonicus*, parasitizes eggs of stink bugs, but no other life stage).

This large parasitoid wasp, relative to the size of the early instar armyworm, is a koinobiont parasitoid. The larva will not kill the caterpillar immediately, allowing it to develop and provide more food for the larva.
Photo: Jack Kelly Clark, UC IPM

Parasitoids are important natural enemies of pest insects. Parasitoids utilize a range of strategies to find their hosts, get their eggs in or on hosts, feed and develop within hosts, and to overcome host defenses against parasitoids. Because parasitoids develop in or on their host, most parasitoids tend to be specialized, attacking one species of host or a group of related species. This specialization makes them highly effective against their host. Parasitoid life cycles are synchronized with that of their host insect and the target life stage they use (ex. adult parasitoids of eggs are active when host insects are producing eggs). Parasitoids can be highly efficient at finding their hosts. Adult female parasitoids often use host–related chemical cues to help them find prey, even when pest population densities are low. In some cases, the host insect gives off chemicals that attract insect hosts, in others, plants fed on by herbivores produce chemicals that attract parasitoids.

Parasitoids have diverse life history characteristics. Two general life history strategies classify parasitoids as idiobionts or koinobionts. **Idiobionts** are characterized as stopping the development of its host by injecting it with a toxin and then laying its egg. The larva of the parasitoid develops in the host that can no longer develop. Idiobionts usually attack a wide range of host insects, and often those in concealed locations. Whereas **koinobionts** parasitize their hosts, usually those in exposed locations, and the host continues to develop with the parasitoid inside of it. Koinobiont parasitoids usually have slowed development and may feed on non-vital tissues first so that the host is not killed immediately, allowing their host to grow larger, sometimes even molting to the next instar or life stage. Because koinobiont parasitoids co-develop with their hosts, they must be able to overcome the hosts' immune defense mechanisms (ex. encapsulation where the host insects blood cells surround and suffocate parasitoid eggs in their body). Therefore, koinobionts often have a narrower, specialized host range.

An emerald ash borer larva is shown with several *Spathius agrili* parasitoid larvae feeding on and eventually killing it. This introduce parasitoid is a gregarious (multiple wasp larvae per host) ectoparasitoid (develops outside the host's body).
Photo: USDA APHIS PPS

Most parasitoids lay their eggs either inside or on the outside of the host's body. **Endoparasitoids** insert their eggs inside the body of their host where the parasitoid larvae develop and feed inside the host's body cavity. Alternatively, **ectoparasitoids** lay their eggs on the exterior surface of the host, where the egg hatches and the larvae attach itself to the outside of the host and feeds on the host through its exoskeleton (skin). Both strategies result in the death of the host insect. Parasitoids can be characterized as solitary or gregarious. In **solitary parasitoids**, a female lays one egg in a host resulting in a single wasp developing and emerging from that individual host. **Gregarious parasitism** is when many (from two to thousands) larvae develop and emerge from one host insect. Gregarious parasitism may result from numerous adult female parasitoids laying eggs in the same host insect, or one adult parasitoid laying multiple eggs into a single host insect.

Parasitoids are beneficial insects providing biological control of pest insects in our managed and natural environments. Their diverse biologies and life histories help them to be successful biological control agents to a wide range of insect taxa.

Weed of the Week

By: Dr. Dan Buonaiuto

I just harvested the first tasty cucumbers from my vegetable patch a few days ago, but you won't want to snack on the spikey fruits this week's featured weed—the wild cousin of my garden cucs—burcucumber (*Sicyos angulatus)*. While burcucumber is native to the eastern United States and should not be controlled in a natural setting, it is known as a tenacious agronomic weed that can significantly compromise corn and soybean yield. Additionally, the rapid growth of this sprawling annual vine can make it problematic in the landscape, nursery or turfgrass context (Figure 1).

Burcucumber Identification:

- Habit: An annual vine that can grow up to 10-20 feet long.
- Leaves: Alternately arranged, wide with 3-5 pointed lobes. Finely pubescent, especially on the underside, most often along the lower veins. Looks similar to cultivated cucumbers (Figure 2).
- Stems: Fuzzy, with clasping tendrils that allow them to climb on most anything.
- Flowers: Plants are monoecious (containing separate male and female flower clusters on the same individual) and are greenish-white to pale yellow in color.

- Fruit: The most definite feature of this species, clusters of small, pointed fruits covered in stiff bristles (Figure 2). Fruits contain a single seed.

Burcucumber Ecology:
Burcumber's preferred habitats include low-lying areas near creeks or rivers, but can also be found in upland locations. This annual plant can produce up to 42,000 seeds per plant and accumulate up to 92 lbs of freshweight in one growing season. Burcumcumber has an extended seed germination window with seeds germinating sporadically from May to August.

Burcucumber Management:
Much of the information about burcucumber control has been studied in an agronomic context. Because this vine is an annual, it spreads exclusively through seed. Therefore, it is important to not let this plant go to seed, which can be achieved through repeated mowing or hand pulling small infestations. Glyphosate, mesotrione and halosulfuron are effective chemical control options in a landscape context. Because seeds germinate sporadically (often after rain events) repeated scouting and treatment is likely necessary over the course of a growing season.

Figure 1: A burcucumber infestation.
Photo: Ohio State Weed Lab, The Ohio State University, Bugwood.org

Figure 2: Foliage and fruits of burcucumber.
Photo: Theodore Webster, USDA Agricultural Research Service, Bugwood.org

Plant of the Week
By: Ginny Rosenkranz

Hydrangea paniculata 'Renhy' Vanilla Strawberry was created in France to have a hydrangea that bloomed early in the season with red flowers. It was named Vanilla Strawberry because the flowers start out pure white and as they mature turn pink and then ruby red in late summer as the temperatures cool down. These plants thrive in the full sun to partial shade, and prefer to grow in organically rich, moist soils. These are the most winter hardy of all the hydrangea, thriving from USDA zones 3-8, and very tolerant of urban conditions. The plants grow 6-8 feet tall and 4-5 feet wide, making them a wonderful specimen plant but also can be used as a summer hedge. Like all the panicle hydrangea, these beautiful plants can be trimmed to the ground late fall or early spring as they bloom on new growth. They can also be pruned to a single stemmed small tree as an accent plant. If left as a shrub, the plants should be trimmed to have only 5-10 primary shoots to create the largest flowers. The foliage is dark green, oval in shape and attach to the stems in an opposite fashion. The terminal flowers are upright, sharply pointed, looking like an ice cream cone and are called panicles. The panicles can grow to 7 inches long and contain both fertile (non-showy) and mostly sterile (very showy) flowers, blooming from July to September

with new flowers forming through the late summer creating vanilla white and pink to ruby red flowers into the cooler days of autumn. The weight of the flowers usually causes the branches to arch gracefully downward. Pest can include some susceptibility to bacteria wilt, bud blight, leaf spot, mildew and rust, while aphids and mites can be occasional visitors.

As the flowers of *Hydrangea paniculata* 'Renhy' Vanilla Strawberry mature later in the season, they turn pink and then ruby red.
Photos: Ginny Rosenkranz, UME

Pest Predictive Calendar “Predictions"

By: Nancy Harding and Paula Shrewsbury

In the Maryland area, the accumulated growing degree days (**DD**) this week range from about **1654 DD** (Clarksville) to **2343 DD** (Nat'l Arboretum/Reagan Nat'l). The Pest Predictive Calendar tells us when susceptible stages of pest insects are active based on their DD. Therefore, this week you should be monitoring for the following pests. The estimated start degree days of the targeted life stage are in parentheses.

Green June beetle – adult emergence **(1539 DD)**
Scarlett oak slug sawfly – larva, early instar **(1544 DD)**
Pine needle scale – egg hatch / crawler (2nd gen) **(1561 DD)**
White prunicola scale – egg hatch / crawler (2nd gen) **(1637 DD)**
Obscure scale – egg hatch / crawler **(1774 DD)**
Orangestriped oakworm – egg hatch / early instar **(1917 DD)**
Magnolia scale – crawler **(1938 DD)**
Fall webworm – egg hatch / early instar (2nd gen) **(1962 DD)**
Maskell scale – egg hatch / crawler (2nd gen) **(2035 DD)**
Euonymus scale – egg hatch / crawler (2nd gen) **(2235 DD)**
Mimosa webworm – larva, early instar (2nd gen) **(2260 DD)**
Japanese maple scale – egg hatch / crawler (2nd gen) **(2508 DD)**

See the Pest Predictive Calendar for more information on DD and plant phenological indicators (PPI) to help you better monitor and manage these pests.

Degree Days (as of July 15, 2026)

Annapolis Naval Academy (KNAK)	2038
Baltimore, MD (KBWI)	2056
Belcamp (FS836)	1956
Clarksville (001MD)	1654
College Park (KCGS)	2121
Dulles Airport (KIAD)	2070
Ft. Belvoir, VA (KDA)	2156
Frederick (KFDK)	1871
Gaithersburg (KGAI)	1985
Greater Cumberland Reg (KCBE)	1842
Martinsburg, WV (KMRB)	1912
Millersville (MD026)	2048
Natl Arboretum/Reagan Natl (KDCA)	2343
Perry Hall (C0608)	1904
Salisbury/Ocean City (KSBY)	2048
St. Mary’s City (Patuxent NRB KNHK)	2191
Westminster (KDMW)	2248

Important Note: We are using the Online Phenology and Degree-Day Models site. Use the following information to calculate GDD for your site: Select your location from the map Model Category: All models Select Degree-day calculatorThresholds in: Fahrenheit °F Lower: 50 Upper: 95 Calculation type: simple average/growing dds Start: Jan 1

Conferences

IPM Diagnostic Session (1 - 3 p.m.)
July 22, 2026
Location: CMREC, Ellicott City, MD

The Stanton A. Gill Symposium: A focus on biological control
August 4 and 5, 2026
Location: CMREC, Ellicott City, MD

FALCAN Truck & Trailer Safety Seminar
October 14, 2026
Location: Urbana Fire Hall, Urbana, MD
Registration opens in July.

Last year's FALCAN Truck & Trailer Safety Seminar sold out well before the event, and we expect another strong turnout this year. Whether you manage a fleet, operate trucks and trailers, or are responsible for compliance and safety, this seminar provides valuable information on the laws, regulations, and best practices that can help protect your business and avoid costly violations. We encourage you to share this information with supervisors, coworkers, and others within your organization who would benefit from attending. Seating is limited, and early registration is recommended.

2027 Advanced Landscape IPM PHC Short Course
This is a recertification short course for arborists, landscapers, IPM consultants, horticulturalists, professional gardeners, and others responsible for urban plant management. The course lectures will be held over four days at the University of Maryland, College Park, MD. In addition, there will be a hands-on lab following lecture (available to a limited number of course attendees). Coordinators: Drs. Paula Shrewsbury and Mike Raupp, Dept. of Entomology, University of Maryland

Lecture dates: Monday, January 4 - Thursday, January 7, 2027 from 8:00 am – 3:00 pm
Lab dates: Monday, January 4 - Thursday, January 7, 2027 from 3:30 pm – 5:30 pm (space limited)
Course information: https://landscapeipmphc.weebly.com/
Registration: https://www.eventbrite.com/e/2027-advanced-landscape-ipm-phc-short-course-tickets-1992372702386?aff=oddtdtcreator

Questions contact: Amy Yaich, 301-405-3911, umdentomology@umd.edu

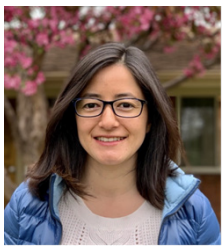
Commercial Ornamental IPM Information
http://extension.umd.edu/ipm

CONTRIBUTORS:

Paula Shrewsbury
Extension Specialist
pshrewsb@umd.edu

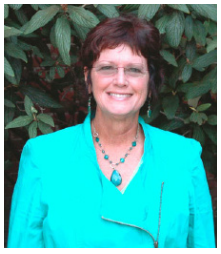
Laura Nixon
Extension Specialist
lnixon1@umd.edu

David Clement
Plant Pathologist
clement@umd.edu



Ana Cristina Fulladolsa
Plant Pathologist
acfulla@umd.edu

Nathan Glenn
Extension Educator
Howard County
nglenn@umd.edu



Nancy Harding
Faculty Research
Assistant

Kelly Nichols
Extension Educator
Montgomery County
kellyn@umd.edu

Dan Buonaiuto
Assistant Professor
Dept. of Plant
Sciences and Land-
scape Architecture
dbuona@umd.edu

Andrew Ristvey
Extension Specialist
aristvey@umd.edu

Ginny Rosenkranz
Extension Educator
Wicomico,
Worcester, Somerset
Counties
rosnkrnz@umd.edu

Thank you to the Maryland Arborist Association, the Maryland Nursery, Landscape, and Greenhouse Association, Professional Grounds Management Society, FALCAN, and USDA NIFA EIP Award # 2024700043556 for their financial support in making these weekly reports possible.