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Beneficial of the Week:

Jumping spiders

Weed of the Week: Wild parsnip (*Pastinaca sativa*)

Plant of the Week: Dwarf crested iris (*Iris cristata*)

Pest Predictive Calendar

Degree Days

Conferences

**Integrated Pest Management
for Commercial Horticulture**
extension.umd.edu/ipm

If you work for a commercial horticultural business in the area, you can report insect, disease, weed or cultural plant problems (**include location and insect stage**) found in the landscape or nursery to sklick@umd.edu

Coordinator Weekly IPM Report:

Laura Nixon, IPM Specialist in Entomology, University of Maryland Extension, Lnixon1@umd.edu, and Paula Shrewsbury, Professor and Extension Specialist in Ornamental and Turf IPM, Department of Entomology, pshrewsbury@umd.edu

Regular Contributors:

Pest and Beneficial Insect Information: Laura Nixon and Paula Shrewsbury (Extension Specialists) and Nancy Harding, Faculty Research Assistant
Disease Information: David Clement (Extension Specialist) and Ana Cristina Fulladolsa (Plant Pathologist and Director, UMD Diagnostic Lab)
Weed of the Week: Kelly Nichols and Nathan Glenn, (UME Extension Educators) and Dan Buonaiuto, (Assistant Professor), Dept. of Plant Sciences and Architecture
Cultural Information: Ginny Rosenkranz (Extension Educator, Wicomico/Worcester/Somerset Counties)
Fertility Management: Andrew Ristvey (Extension Specialist, Wye Research & Education Center)
Design, Layout and Editing: Suzanne Klick (Technician, CMREC)

Spotted Lanternfly Eggs Continue to Hatch in Lots of Locations

By: Paula Shrewsbury

Last week we had the first reports of spotted lanternfly (SLF, *Lycorma delicatula*) eggs hatching in MD. This week there are numerous reports of SLF eggs hatching. This week first instar nymphs of SLF have been found in MD in: Centerville on maple by Kevin Schroeder (Davey Tree); Gaithersburg by Heather Zindash (The Soulful Gardener); Washington DC on maple by Sam Fisher (Bartlett Tree Experts); College Park on river birch by Alec Lynde; and Georgetown (Wash. DC) on Virginia Creeper by Marc Vedder. Thanks to all of you who have kept us informed of what you are seeing in the field!

We should continue to see more SLF eggs hatching this week. If you look at the degree day (DD) accumulations listed at the end of the newsletter you will see that most locations have reached or surpassed 270 DD, the number of degree days that let us know it's time to monitor for SLF egg hatch. [Click here to see a time lapse video of SLF nymphs wriggling their way of the eggs \(video by M.J. Raupp, UMD\).](#)

What should you do with all the SLF nymphs that will be on plants soon? It depends on the circumstances. In landscapes, at this time of year when SLF are in early nymphal stages on established trees and other vegetation you likely do not need to do much. The nymphs are small and do not produce much honeydew and they move a lot among different types of plants and

vegetation. *In landscapes and nurseries*, if you are moving plants or other materials with SLF nymphs, then you need to follow [state quarantine and treatment regulations](#) which involves removing / getting rid of SLF on plants before movement. [For information on chemical and other control options for SLF, and their biology and ecology, go to this Penn State Extension SLF Management Guide](#). Be sure to consider pollinator and natural enemy protection when using chemical options.



Newly hatched spotted lanternfly nymphs. The white nymphs were less than a few hours old and the black and white nymphs were just a little bit older.

Photo: Kevin Schroeder, Davey Tree



Throughout the DMV, spotted lanternfly nymphs are popping the lid (operculum) at the top of their egg case and emerging head-first from eggs deposited last autumn by females. The emerging nymphs are white with distinct black eyes for about an hour or so.

Photo: M.J. Raupp, UMD

Box Tree Moth Update

We continue to monitor box tree moth caterpillar activity at the same sites in Washington County, MD this week. We found 3rd and 4th instar caterpillars, and several possible 5th instars. There is still no sign of pupae or adults. If you see any BTM activity, please report it to the IPM Alerts and your state department of agriculture.



Box tree moth caterpillars are active in Washington County this week. Photo: Suzanne Klick

Cold Damage

By: Suzanne Klick

We received multiple reports of damage to plants when temperatures throughout the area dropped into the 20s (°F) on the morning of April 21. The cold temperatures damaged a wide variety of plants including a big leaf magnolia and crape myrtles in Silver Spring (Amanda Laudwein), Redbud 'Merlot', *Taxodium*, and honeylocust in Frederick County (Marie Rojas), and tulip tree in Columbia (Bill Stocker). Marie also saw a report online of damage to concolor firs with drooping young candles and extensive damage to grapes in Frederick County. Be sure to monitor damaged plants closely. Damaged plant tissue can provide entry points for fungi and bacterial disease pathogens.



Redbud 'Merlot' (left photo) and *Taxodium* (photo above) with significant freeze damage.
Photos: Marie Rojas, IPM Scout



Heavy freeze damage on this big leaf magnolia.
Photo: Amanda Laudwein



Some of the leaves on this tulip tree were damaged by the freezing temperatures.
Photo: Bill Stocker

Viburnum Leaf Beetle Larvae – Busy skeletonizing leaves of viburnum

By: Paula Shrewsbury

On Monday April 20th, Mike Raupp (UMD) reported viburnum leaf beetle, *Pyrrhalta viburni* (Chrysomelidae) larvae were active causing heavy skeletonization on viburnum foliage at Lake Roland in Baltimore County MD. Miri Talabac (HGIC UME) also reported active viburnum leaf beetle larvae this week.

Viburnum leaf beetle is an introduced pest that was first detected in the U.S. in upstate NY in 1996 and in 2017 in Washington County MD. It has increased its range and is currently found in Maryland, New York, Maine, northern Pennsylvania, Vermont, parts of Ohio, Minnesota, Michigan, Canadian Maritime provinces, Ontario, and British Columbia (and possibly other locations).

Viburnum leaf beetles have one generation a year, overwinter as eggs inserted in stems, and eggs hatch around 210 DD. Most areas of MD have reached and/or passed the 210 DD threshold for egg hatch. If you manage viburnums, you should monitor closely for feeding damage by larvae which starts as skeletonization and can progress to complete defoliation. Larvae pupate in the soil around mid-June, and adults emerge and are active from July through September. Adults also feed on viburnum foliage leaving oblong holes in leaves. In late summer and fall, females chew small holes in twigs and lay eggs in these cavities where they spend the winter. High populations can completely defoliate a shrub, sometimes resulting in shrub death.



Viburnum leaf beetle larva and feeding damage on viburnum.
Photo: Miri Talabac, HGIC UME



Adult viburnum leaf beetle.
Photo: Kent Loeffler from Cornell University

Recommendations. If viburnum leaf beetle larvae are currently damaging plants, consider treating with a spinosad based product, some of which are reduced risk or organic products, or another labelled product. When eggs are present on twigs of viburnums, prune out and destroy the twigs to reduce populations in the spring.

Research out of Cornell University has demonstrated that viburnums vary in their susceptibility to viburnum leaf beetle. Click here to see lists of susceptible and resistant viburnum species - <http://www.hort.cornell.edu/vlb/suscept.html>. This information will inform you on which plants to focus monitoring, pest management decision making, and selection choices for which viburnum species to include in landscapes or not.

For more detailed information and pictures of viburnum leaf beetle go to:

<https://extension.umd.edu/resource/viburnum-leaf-beetle/>

<http://www.hort.cornell.edu/vlb/>



Defoliation caused by viburnum leaf beetle adults.

Photo: Paul Weston, Cornell University, Bugwood.org

Elder Shoot Borer Activity Has Started

Marie Rojas, IPM Scout, found elder shoot borers, *Achatodes zae* (Noctuidae), inside *Sambucus nigra* 'Eva', causing flagged tips. Recently hatched caterpillars are boring into new shoots. When they reach maturity in early summer, they tunnel into dead stalks to pupate. Look for frass at the base of old wood. Prune out infested green shoots or mature canes during the growing season. In winter, remove dead canes to reduce pupation habitat. Be sure to remove pruned cuttings from the area.



Elder shoot borer caterpillar found in a flagged tip of *Sambucus nigra*.
Photo: Marie Rojas, IPM Scout

Roseslug Sawfly

By: Suzanne Klick

Miri Talabac, HGIC, found roseslug sawfly larvae (*Endelomyia aethiops*) feeding on Knock Out® roses this week. Look for the small larvae on the undersides of foliage. The last instar reaches about 1/2 long. It will drop to the ground, make a cocoon, and remain in the soil until they pupate the following spring. This species will feed until late May or early June in Maryland. There is only one generation. The other two common roseslug species (bristly roseslug and curled roseslug sawflies have multiple generations and can cause damage throughout the late spring and summer).

Monitor roses early in the season. The larvae feed for about a month depending on temperatures and weather patterns. If the population is low, consider hand picking the larvae or spraying plants with a hose to manage this pest. Horticultural oil or insecticidal soap are contact insecticides that are effective, but be sure to get full coverage of the larvae. Spinosad (e.g. Conserve) is another option with low impact on beneficials.



Roseslug sawfly larvae (*Endelomyia aethiops*) cause etching damage in leaves. Eventually, these areas will turn brown and drop to the ground leaving holes in the foliage.

Photos: Miri Talabac, UME-HGIC

Spiny Ash Sawfly on Fringe Tree

By: Nancy Harding and Paula Shrewsbury, UMD

This week in Bowie, my white fringe tree (*Chionanthus virginicus* (Family: Oleaceae) was in full bloom, looking amazing with its fragrant, white, fringe-like flowers. However, I noticed a lot of defoliation and shot holes of varying sizes on the foliage. The culprit who is munching on the foliage is the **spiny ash sawfly**, *Eupareophora parca* (Hymenoptera: Tenthredinidae). Young larvae are light green and late instar larvae have a wide dark band along their top (dorsal) side with yellow along the sides. All larval stages have spines which become darker and more pronounced in the later larval stages.

The spiny ash sawfly is known to occur in two ranges, in the eastern United States and southeastern Canada, and in coastal California and Oregon. It has one generation per year and overwinters as pupae in the ground. Larvae are active from around late March through early July. *Eupareophora* is known to feed on *Fraxinus americana* (white ash), *Fraxinus nigra* (black ash), *Fraxinus oregona* (Oregon ash), *Fraxinus*

pensylvanicus (green ash), *Carya illinoensis* (pecan), and *Chionanthus* (fringe tree). Fringe tree are in the same family as ash (Oleaceae) so it is not surprising to see this sawfly feeding on fringe tree too. Spiny ash sawfly has been reported as a pest on ash in some urban locations.

Control: If densities of spiny ash sawfly are relatively low and reachable, manually remove them by handpicking, shaking branches, or using a strong blast of water, as they cannot easily crawl back up the tree. For larger infestations, applying insecticidal soap, neem oil, or spinosad directly to larvae, or another labeled product, should reduce populations.

For more information and images of damage and different life stages of *Eupareophora parca* go to:

<https://www.inaturalist.org/taxa/495096-Eupareophora-parca>

https://www.jungledragon.com/specie/24472/spiny_ash_sawfly.html



Early instar spiny ash sawfly, *Eupariophora parca*, and feeding damage on white fringe tree.
Photos: N. Harding, UMD



Late instar spiny ash sawfly, *Eupariophora parca*, larvae.
Photo: M. Lankford (CC By-NC), iNaturalist

Pearleaf Blister Mites

By: Paula Shrewsbury

Marie Rojas, IPM Scout, found pearleaf blister mites, *Eriophyes pyri* (Eriophyidae), just starting to feed on the leaves of *Pyrus shinsui* in Frederick County MD. This eriophyid mite is found on edible and ornamental pears, *Pyrus* sp. and feeds on newly emerged leaves in the spring. Timing of treatment is best done prior to their movement into the leaf blisters after petal fall. A 2% horticultural oil application is a good control option. Management is often timed for the fall to treat the mites while they are in the outer bud scales. Delayed dormant applications also target the bud scales with oils.



Early damage to *Pyrus shinsui* by pearleaf blister mite.
Photo: Marie Rojas, IPM Scout

Eastern Tent Caterpillar Update

By: Paula Shrewsbury

Eastern tent caterpillar (ETC), *Malacosoma americana* (Lepidoptera: Lasiocampidae) started hatching about a month or so ago. At this time the tents, and the caterpillars in them, are quite large. Very soon the caterpillars will leave their protective tents to find a place to pupate on the ground. If caterpillars are still causing damage and/or a nuisance, the best response is to get a big stick and rip open the tents exposing the caterpillars to predators. In the winter, before March of next year, remove any egg masses that the female moths may lay on the small branches of their host trees.



A large Eastern tent caterpillar tent with caterpillars.
Tents are formed in tree forks.
Photo: P.M. Shrewsbury, UMD

Which Psyllid Species is on Your Boxwood?

By: Paula Shrewsbury and Nancy Harding, UMD

Since the article we wrote about a newly re-discovered psyllid on boxwoods in the [April 10, 2026 IPM Report](#), we have received several emails from people pretty sure they have the re-discovered *Spanioneura fonscolombii* rather than *S. buxi*. The best way to diagnose which psyllid species you have is to catch an adult and look for the 4 dark spots on the front edge of each forewing which indicates it is *S. fonscolombii*. In addition, *S. fonscolombii* tend to produce a lot more of the white wax and the new leaves are not cupped like with *S. buxi*. Go back to the April 10 IPM Report to review the images and other characteristics to tell them apart.



Abundant wax, likely produced from *Spanioneura fonscolombii* on boxwood.

Photo: Michael Garner, Brightview Landscape Services

Rabbit Damage

Elaine Menegon, Good's Tree and Lawn Care, found rabbit damage on an arborvitae in Hershey, PA this week. The winter snow provided cover for rabbits to travel farther up the tree than usual and cause significant damage at the base.



Rabbit damage at the base of this arborvitae.
Photo: Elaine Menegon, Good's Tree and Lawn Care

Cutworms – Heavy damage to lawn turf

By: Paula Shrewsbury

On Thursday, Kevin Bohrer (Goshen Enterprises, Inc), sent in images of severe damage to turfgrass in a residential lawn in Laytonsville, MD on April 23rd. Damage to the lawn (mainly fescue) on the right started last week, and the lawn damage on the left started this week (see image). Kevin said he and other turf professionals he consulted have seen this damage at several locations in the Laytonsville and Damascus areas. He found what looks like early and mid-life stage **bronzed cutworm, *Nephelodes minians* (Noctuidae) caterpillars** in the turf (see image). It is difficult for me to provide a positive “for sure” diagnosis as to what is causing the damage based on pictures. We do know bronzed cutworms can be pests of lawn turf in residential settings and feed on a range of turf types. After further communication with Kevin and his description of the damage symptoms he saw, and the conversations he had with two other turf professionals, it sounds like the culprit of the damage is very likely bronzed cutworm caterpillars.

Bronzed cutworm usually overwinter as eggs and the larvae hatch early in the spring with most damage happening in the spring / early summer. There is only one generation per year. Bronzed cutworm caterpillars feed down into the thatch and crowns resulting in circular holes/ burrows and when populations are high, they can consume large amounts of grass resulting in a thinning appearance to the turf. Look for signs of cutworm which include cut leaf blades, piles of greenish fecal pellets, and the presence of larvae in the thatch. The best way to monitor these nocturnal cutworms during the day is to use an irritant flush (mixture of about 1-2 tablespoons of dish soap to a gallon of water) by pouring the solution over a few square feet of turf. The soap will cause the caterpillars to move to the surface within about 5 minutes and then they are easier to count. Birds foraging in the soil can also indicate a caterpillar or other insect pest is present.

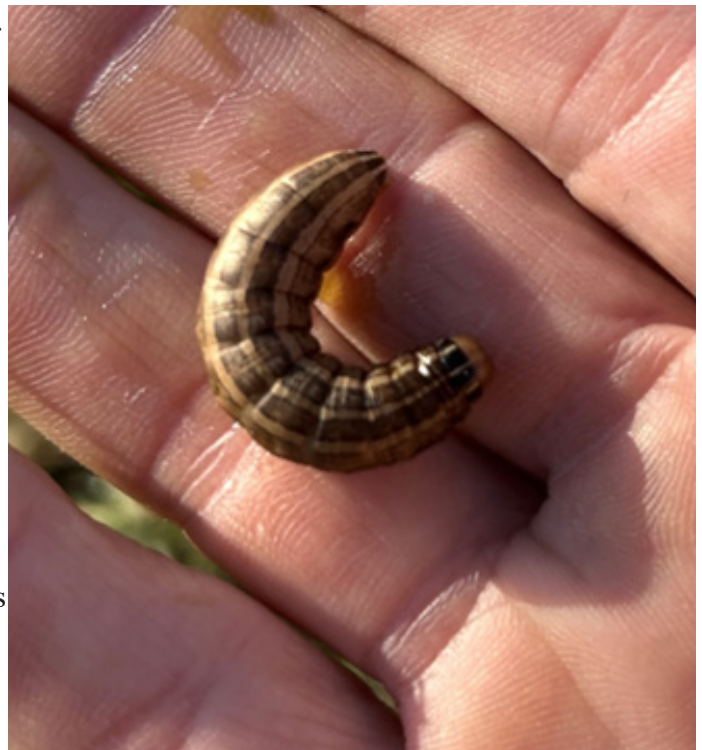
Damage tends to be more severe on stressed turf and turf that is mowed too low. With the severe drought we are having in this area it is likely the turf is stressed. A wide range of products are labeled for cutworm management including biologicals such as Bt products and entomopathogenic nematodes, and Spinosad, in addition to numerous other pesticides ([click here for a partial list of pesticides](#)).

If you are seeing damage to turf and signs of cutworms, please let us know (pshrewsbury@umd.edu; sklick@umd.edu).



Damaged lawn turf in Laytonsville, MD, which is likely caused by bronzed cutworm. Damage to the lawn on the right started last week, and the lawn damage on the left started this week.

Photo: Kevin Bohrer, Goshen Enterprises, Inc.



Bronzed cutworm found in damaged lawn turf.

Photo: Kevin Bohrer, Goshen Enterprises, Inc.

Ambrosia Beetle Update

By: Paula Shrewsbury

We continue to monitor ambrosia beetle activity in traps located at the Central MD Research and Education Center (CMREC) in Ellicott City, Beallsville and Gaithersburg, MD (traps monitored by Marie Rojas, IPM scout), and Salisbury, MD (trap monitored by Ginny Rosenkranz, UME). We also receive reports from people in the field (you guys) on trap catches and adult boring activity in trees. We are monitoring closely for the following key pest ambrosia beetles: *Xylosandrus germanus* (**black twig borer**), *X. crassiusculus* (**granulate ambrosia beetle**), and *Cnestus mutilatus* (**camphor shot borer**), and keeping an eye out for other potential troublemakers. Please let us know if you have ambrosia beetle activity on trees (sklick@umd.edu).

Symptoms and signs of ambrosia beetles are poor plant growth, wet areas on tree trunks from tree sap, frass tubes (toothpicks) produced by beetles boring, and small circular entry holes that may or may not have adult beetles or sawdust in or around them. The *optimal conditions* for ambrosia beetle flight are warm temperatures >70°F for 2-3 consecutive days, especially following periods of rain. Ambrosia beetles have a broad range of host tree species that they may attack. I suggest monitoring of known susceptible trees of *Xylosandrus* species such as styrax, yellowwood, birch, zelkova, and redbud. Other hosts reported to be attacked include the hybrid series of *Cornus florida* and *C. kousa*, Kwanzan cherries, *Ilex opaca* 'Satyr Hill' and 'Miss Helen', and paperback maples. Last week we had reports by Marie Rojas (IPM Scout) of *adult activity* (frass “toothpicks” from tree trunks) in American holly (*Ilex opaca*) and dogwood (*Cornus* 'Appalachian Spring') in Beallsville, MD on April 17th; and activity in a nursery in Monkton, MD on Kousa dogwood, Kwanzan cherry, and crabapple on April 13th. We have received no other reports of ambrosia beetles in trees this week. This week our trap at CMREC in Ellicott City MD caught about 25 ambrosia beetles. We have not been able to identify which species they are yet.

Recommendations:

The best IPM strategy for Ambrosia beetle is to implement practices to avoid and reduce tree stress. If you have a pre-existing stressor in some of your trees, such as pests, diseases, or areas that may be under drought or are “too wet”, be sure to closely monitor those areas. Monitoring can consist of traps (see the [March 27, 2026 IPM Report](#) article on this) and visual monitoring for signs and symptoms of adult and larval activity.

When ambrosia beetle adults are active, permethrin and bifenthrin products are registered for trunk sprays in nurseries and landscape and have been found through research to give good control if timed accurately. Be sure to check product labels and follow pollinator protection recommendations. The



Ambrosia beetle frass “toothpicks” resulting from adult beetles boring into this *Cornus* ‘Appalachian Spring’. Damage symptoms were found on April 17, 2026 in Beallsville, MD.

Photo: Marie Rojas, IPM Scout



Body of ambrosia beetle adult at the entrance of its boring hole.

Photo: Paula Shrewsbury, UMD

idea is to protect the tree trunks with an insecticide barrier so when adults chew their way through the bark, they become poisoned and die. Once the beetle adults and larvae are in the tree, there is not a lot that can be done. If beetle densities are high in infested trees, remove and destroy the tree(s) to reduce the beetle source and movement to other trees.

The weather this week has been variable in temperature and rain. At the beginning of the week temperatures were hitting greater than the ~70 °F threshold for ambrosia beetle adult activity, and then the temperatures are cooling down for the next several days starting later today. *Given this and the reported activity in traps and trees in recent weeks, be sure to monitor judiciously. I strongly recommend that you consider treating susceptible trees to prevent adults from successfully boring into the trees and laying eggs.*



Sawdust around the entrance hole made an ambrosia beetle boring into a tree trunk.
Photo: Paula Shrewsbury, UMD

Powdery Mildew on Kentucky Bluegrass Sod

By: Suzanne Klick

Mark Schlossberg, ProLawn Plus, Inc., found powdery mildew infecting Kentucky bluegrass sod in a shady area in Stevenson, MD this week. Poor air circulation, high humidity, and cool temperatures promote powdery mildew infection in turf areas. It's usually not a problem in turf and infection is reduced as temperatures increase. Increased sunlight (if pruning trees shading the area is possible), improved airflow, and reduced nitrogen fertilizer can help manage this disease. Fungicides are usually not needed to control powdery mildew in turf.



Powdery mildew is infecting this area of Kentucky bluegrass.
Photo: Mark Schlossberg, ProLawn Plus, Inc.

Beneficial of the Week

By: Paula Shrewsbury

Jumping spiders: beautiful, beneficial and interesting

As I was working on my computer yesterday, I looked over at my monitor and there was a jumping spider (Salticidae) walking across the screen and then it continued to explore the rest of the monitor. After observing it for a few minutes, I caught the spider and put it outside where it should hopefully live a better life. When observing plants outside, it is not unusual to find jumping spiders (family Salticidae) of all sizes and colors. Jumping spiders' range in size from 3 to 12 mm (0.12 to 0.5") and vary in colors that camouflage against tree bark to bright metallic colors. The diversity and number of jumping spiders that are active at any given time is quite impressive and makes me appreciate the biological control they provide throughout the season.

Salticidae is the largest family of spiders with over 6,000 species of jumping spiders worldwide and more than 300 species found in the U.S. These spiders gain their name because not only do they crawl and run quickly but they often move by making short jumps from one location to another, jumping as far as 10 – 50 times their body length. Jumping spiders are recognized by their somewhat rectangular or box-like cephalothorax (the head and thorax areas that are fused together in spiders compared to insects) which has 1 row of 4 very distinct dark eyes along the front facing edge of the cephalothorax with the center 2 eyes being quite large compared to the others, and 2 more pairs of eyes along the top facing edge of the cephalothorax (=8 eyes total).

Jumping spiders have some very interesting behaviors. Male jumping spiders differ from females in that they may have plumose, colored or iridescent hairs, front leg fringes, structures on other legs, and other modifications used in "courting" females. In many species the male performs complex courtship displays or "dances" in which he moves his body up and down or in zig zag patterns and waves his front legs in a highly specific manner to impress a female. There are also specific sound effects (ex. drumming on a substrate such as a plant with his legs or abdomen) that the male makes associated with this courtship behavior. If the female "likes" or accepts the male they will then mate. After



Close up of an Emerald jumping spider showing its diagnostic front row of eyes with two large eyes in the middle and a smaller eye on either side of the large (plus the 4 other small eyes behind this row). Also note the metallic green colors.

Photo: Jim Moore, Maryland Biodiversity Project



A jumping spider enjoying a newly captured caterpillar meal.

Photo: Michael Raupp, UMD

mating, the female lays her eggs in shelters (ex. under stones or bark) lined with silk or on the surface of plants. The female will often guard the eggs and the newly hatched spiderlings until they are old enough to forage for food on their own.

To see a fun and entertaining video of some of the interesting male mating behaviors put to the music of “Stayin Alive” by the Bee Gees, go to: https://youtu.be/HPh_Gi7PCqs. It will make you laugh and maybe dance too!



A magnolia green jumper (Salticidae) guarding her newly hatched spiderlings.

Photo: Bob Cammarata, Maryland Biodiversity Project

Jumping spiders are not web builders. They actively hunt for their prey by foraging on the leaves and branches of plants. They usually hunt during daylight and have very good eyesight. Jumping spiders sneak up to within a few body lengths of their prey, crouch, crawl slowly forward, and then lift

their front legs and pounce on their prey to capture it. Once captured the spider immobilizes its prey by injecting them with venom and release proteolytic enzymes into or onto the prey which liquefies the tissue allowing the spider to slurp up its food. To see a salticid spider capturing its caterpillar prey go to: https://youtu.be/py_V2lqWpb4 or another [preying on a mosquito](#). Most jumping spiders are considered to be generalist predators, meaning they will feed on a broad diet of different types of insects. Jumping spiders have been seen eating stink bugs, lace bugs, flies (including mosquitoes), caterpillars, beetles, moths, and other mobile insects. Some salticid species are also known to include nectar as a food resource. Spiders make up a significant part of the natural enemy assemblage in our ornamental and turfgrass systems helping to prevent plant feeding insects from reaching population levels that cause economic or aesthetic damage. Be sure to look for these jumping spiders in landscapes and nurseries you manage and to appreciate the benefits they provide.

Weed of the Week

By: Dan Buonaiuto

Make sure to get your gloves out for this week’s weed of the week: *Pastinaca sativa* (wild parsnip, Figure 1). Sap from this biennial weed contains a phytotoxin that can cause severe blistering to the skin, so control with care. Wild parsnip can be found in both sun and shade, in pastures and agricultural fields, along roadsides or in other disturbed areas. Here in Maryland, we are at the southern edge of this species’ range, but it is found throughout the state.

Wild Parsnip Identification:

- **Growth habit:** This biennial plant forms a rosette and its first year of growth, sending up a flowering stalk up to 5 feet tall in its second year. This floral stalk is hollow, somewhat hairy and has deep vertical grooves.
- **Leaves:** Leaves are compound, with 5-15 serrated leaflets that are attached by a long petiole that encircles the stem. Leaves and stems are yellow-green in color (a natural example of the color chartreuse).
- **Flowers:** As a member of the carrot (Apiaceae) family, wild parsnip has a characteristic umbrella shaped inflorescence (Figure 2). The yellow flowers are small, growing in clusters that are usually more than 4 inches in diameter. Flowering typically occurs in June-July.

Wild Parsnip Ecology:

Wild parsnip was introduced as a food source by early European settlers. This plant is toxic to livestock and degrades bird habitat. Wild parsnip sap contains a chemical called furocoumarin which adheres to unprotected skin. Sunlight activates furocoumarin, causing extreme skin burns (phytophotodermatitis) which can require hospitalization, and affected areas may never fully recover.

Reproduction is through seed, with a single individual able to produce thousands of wind-dispersed seed.

Wild Parsnip Control:

To minimize the risk of phytophotodermatitis, always fully cover skin when working with this species. Many landscape managers also opt to control wild parsnip in the evening and wash the toxins off before exposure to sun the next day.

Control can be achieved with both mechanical and chemical methods, but efficacy of these approaches depends on the life stage of the target plants.

At the rosette stage, applications of glyphosate or triclopyr are useful but will not be effective after the plant has bolted. From bolting until the primary flower head develops, cutting the taproot with a shovel several inches below the surface can be effective. This mechanical approach works earlier in the plant lifecycle as well, though it will likely resprout. There is also a limited time window for mowing, between the development of the primary flower heads and secondary (lateral) flower heads.



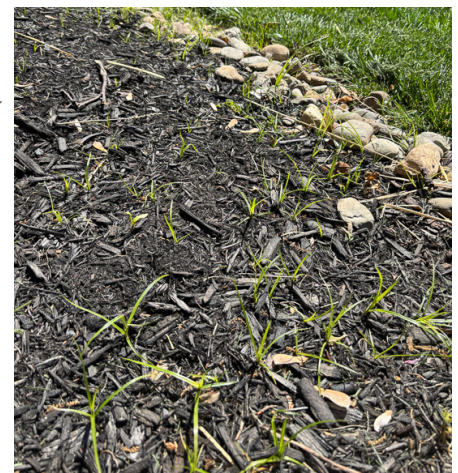
Figure 1: Wild parsnip plants
Photo: Leslie J. Mehrhoff,
University of Connecticut,
Bugwood.org



Figure 2: Wild parsnip flowerhead.
Photo: John Cardina,
The Ohio State
University, Bugwood.
org

Nutsedge is Emerging

Mark Schlossberg, ProLawn Plus, Inc., is reporting that nutsedge is already up in northern Baltimore City on April 21. This perennial weed reproduces by seed, rhizomes, and tubers and is difficult to control.



Difficult to control nutsedge is already up in Baltimore City.
Photo: Mark Schlossberg, ProLawn Plus, Inc.

Plant of the Week

By: Ginny Rosenkranz

Iris cristata, also known as dwarf crested iris, is a native herbaceous perennial that grows only 6-8 inches tall and spreads 6 to 12 inches wide. The plants thrive with morning sun and light afternoon shade and organically rich, moist, well drained soils. Like many irises, the dwarf crested iris spreads by branching rhizomes and can create a lovely dense groundcover. They typically bloom in April with 3 small pale lavender-blue 'standards' which are the petals and 3 larger 'falls' that are the sepals, that are also pale lavender-blue but with a white center blotch and bright yellow crested ridges. The 3-inch, showy flowers are on very short stems. The lance-shaped, bright green leaves can grow up to 6 inches long and have a pale green underside. When they are in bloom, the flowers attract hummingbirds and bumble bees. These lovely spring beauties are cold tolerant in USDA zones 3-9, provide a groundcover in shady areas, and are very deer resistant. Slugs and snails can be problematic as the plants thrive in the same areas that they do. There are a few cultivars including 'Alba' which has pure white flowers with the golden crested ridge, 'Caerulea' with deep dark blue flowers, 'Eco White Angel' that is a bit larger than the native, and 'Shenandoah Sky' that is a pale blue in color.



Dwarf crested iris flowers attract hummingbirds and bumble bees.

Photos: Ginny Rosenkranz, UME

Pest Predictive Calendar “Predictions”

By: Nancy Harding and Paula Shrewsbury

In the Maryland area, the accumulated growing degree days (**DD**) this week range from about **273 DD** (Clarksville) to **456 DD** (Nat'l Arboretum/Reagan Nat'l). The [Pest Predictive Calendar](#) tells us when susceptible stages of pest insects are active based on their DD. Therefore, this week you should be monitoring for the following pests. The estimated start degree days of the targeted life stage are in parentheses.

Roseslug sawfly – larva, early instar (**230 DD**)
Elongate hemlock scale – egg hatch / crawler (1st gen) (**232 DD**)
Boxwood leafminer – adult emergence (**249 DD**)
Hawthorn lace bug – first adult activity (**259 DD**)
Spotted lanternfly – egg hatch (**270 DD**)
Bristly roseslug sawfly – larva, early instar (**284 DD**)
Imported willow leaf beetle – adult emergence (**290 DD**)
Hawthorn leafminer – adult emergence (**292 DD**)
Andromeda lace bug – egg hatch (**305 DD**)
Pine needle scale – egg hatch / crawler (1st gen) (**307 DD**)
Cooley spruce gall adelgid – egg hatch (**308 DD**)
Eastern spruce gall adelgid – egg hatch (**308 DD**)
Spirea aphid – adult / nymph (**326 DD**)
Lilac borer – adult emergence (**350 DD**)
Melon aphid – adult / nymph (**351 DD**)
Spiny ash sawfly – larva, early instar (**358 DD**)
Spongy moth (formerly gypsy moth) – egg hatch (**373 DD**)
Holly leafminer – adult emergence (**375 DD**)
Hemlock woolly adelgid – egg hatch (2nd gen) (**411 DD**)
Basswood lace bug – first adult activity (**415 DD**)
Emerald ash borer – adult emergence (**421 DD**)
Locust leafminer – adult emergence (**429 DD**)
Honeylocust plant bug – egg hatch, early instar (**433 DD**)
Fourlined plant bug – egg hatch, early instar (**435 DD**)
Lesser peachtree borer – adult emergence (1st gen) (**468 DD**)
Oak erricoccin scale (oak felt scale) – egg hatch / crawler (**469 DD**)
Maskell scale – egg hatch / crawler (1st gen) (**470 DD**)
Oystershell scale – egg hatch / crawler (1st gen) (**486 DD**)
Minute cypress scale – egg hatch / crawler (**511 DD**)
White prunicola scale – egg hatch / crawler (1st gen) (**513 DD**)
Euonymus scale – egg hatch / crawler (1st gen) (**522 DD**)
Bronze birch borer – adult emergence (**547 DD**)
Potato leafhopper – adult arrival (**603 DD**)
Black vine weevil – adult emergence (**607 DD**)

See the [Pest Predictive Calendar](#) for more information on DD and plant phenological indicators (PPI) to help you better monitor and manage these pests.

A Lot of Potential Insect Pest Activity – Check out what to expect

By: Paula Shrewsbury and Nancy Harding

It is the time of year when an abundance of insect species are becoming active. Every week near the end of the IPM Report, we list degree day (DD) accumulations for several areas in MD and Washington D.C. for the week. In addition, we go to the [Pest Predictive Calendar](#) and include a list of potential pest insects that may be active in the current range of DD accumulations for that week. We are at the time of year when lots of key pests and other “important pests some years” target stages are becoming active. Go to the end of this report and look at the list. These are insects you should be monitoring for. For example, this week in MD, DD accumulations ranged from 273 DD to 456 DD depending on location. There are several scale species whose crawler stages become active within this range (ex. elongate hemlock scale, maskell scale, white prunicola scale). Don’t forget emerald ash borer adults, adelgids, and many lace bug species also become active. Hopefully, you can use this information to improve your monitoring and pest management programs.



Adult emerald ash borer on ash foliage that recently emerged from under the bark of an ash tree where it spent the winter as a larva and pupa. Note the defoliation of the ash leaf where the beetle had been feeding.

Photo: Leah Bauer, USDA Forest Service Northern Research Station, Bugwood.org



White bundles of fluffy wax on Hemlock are a sure sign of a woolly adelgid infestation.

Photo: Michael Raupp, UMD



Underside of an azalea leaf showing azalea lace bug adults and fecal spots.

Photo: P.M. Shrewsbury, UMD

Degree Days (as of April 22, 2026)

Annapolis Naval Academy (KNAK)	315
Baltimore, MD (KBWI)	365
Belcamp (FS836)	308
Clarksville (001MD)	273
College Park (KCGS)	405
Dulles Airport (KIAD)	413
Ft. Belvoir, VA (KDA)	424
Frederick (KFDK)	323
Gaithersburg (KGAI)	377
Greater Cumberland Reg (KCBE)	367
Martinsburg, WV (KMRB)	376
Millersville (MD026)	384
Natl Arboretum/Reagan Natl (KDCA)	456
Perry Hall (C0608)	316
Salisbury/Ocean City (KSBY)	394
St. Mary's City (Patuxent NRB KNHK)	404
Westminster (KDMW)	446

Important Note: We are using the [Online Phenology and Degree-Day Models](#) site. Use the following information to calculate GDD for your site: Select your location from the map Model Category: All models Select Degree-day calculator Thresholds in: Fahrenheit °F Lower: 50 Upper: 95 Calculation type: simple average/growing dds Start: Jan 1

Conferences

May 20, 2026

[Mid-Atlantic Nursery Workshop: the Water Loop to Pot: Managing Ponds, Irrigation, Substrates, and Runoff for Better Production Profits](#)

Location: Hampton Roads Agricultural Research and Education Center, Virginia Tech, Virginia Beach, VA

June 16, 2026

[2026 Eastern Shore Procrastinators Conference](#)

Location: Zoom

June 18, 2026

MNLGA Field Day

Location: Mt Cuba Center, Hockessin, DE

June 26, 2026

[Montgomery County Pesticide Procrastinators Conference](#)

Location: Derwood, MD

IPM Scouts' Diagnostic Session (1 - 3 p.m.)

June 23, 2026

Location: CMREC, Ellicott City, MD

Commercial Ornamental IPM Information

<http://extension.umd.edu/ipm>

CONTRIBUTORS:



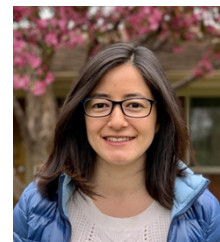
Paula Shrewsbury
Extension Specialist
pshrewsb@umd.edu



Laura Nixon
Extension Specialist
lnixon1@umd.edu



David Clement
Plant Pathologist
clement@umd.edu



Ana Cristina Fulladolsa
Plant Pathologist
acfulla@umd.edu



Nathan Glenn
Extension Educator
Howard County
nglenn@umd.edu



Nancy Harding
Faculty Research
Assistant



Kelly Nichols
Extension Educator
Montgomery County
kellyn@umd.edu



Dan Buonaiuto
Assistant Professor
Dept. of Plant
Sciences and Land-
scape Architecture
dbuona@umd.edu



Andrew Ristvey
Extension Specialist
aristvey@umd.edu



Ginny Rosenkranz
Extension Educator
Wicomico,
Worcester, Somerset
Counties
rosnkranz@umd.edu

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