

In This Issue...

- [Box tree moth update](#)
- [Boxwood spider mites](#)
- [Boxwood leafminer](#)
- [Maskell scale](#)
- [Boxelder bug adults](#)
- [Rose rosette disease](#)
- [Roseslug sawfly](#)
- [Greenhouse/cut flower alert](#)
- [New species of boxwood psyllid](#)

Beneficial of the Week:

Zebra swallowtails

Weed of the Week:

Crabgrass (*Digitaria sanguinalis*)

Plant of the Week: Virginia bluebells (*Mertensia virginica*)

Pest Predictive Calendar

[Degree Days](#)

[Conferences](#)

Integrated Pest Management for Commercial Horticulture
extension.umd.edu/ipm

If you work for a commercial horticultural business in the area, you can report insect, disease, weed or cultural plant problems (**include location and insect stage**) found in the landscape or nursery to sklick@umd.edu

Coordinator Weekly IPM Report:

Laura Nixon, IPM Specialist in Entomology, University of Maryland Extension, Lnixon1@umd.edu, and Paula Shrewsbury, Professor and Extension Specialist in Ornamental and Turf IPM, Department of Entomology, pshrewsbury@umd.edu

Regular Contributors:

Pest and Beneficial Insect Information: Laura Nixon and Paula Shrewsbury (Extension Specialists) and Nancy Harding, Faculty Research Assistant
Disease Information: David Clement (Extension Specialist) and Ana Cristina Fulladolsa (Plant Pathologist and Director, UMD Diagnostic Lab)
Weed of the Week: Kelly Nichols and Nathan Glenn, (UME Extension Educators) and Dan Buonaiuto, (Assistant Professor), Dept. of Plant Sciences and Architecture
Cultural Information: Ginny Rosenkranz (Extension Educator, Wicomico/Worcester/Somerset Counties)
Fertility Management: Andrew Ristvey (Extension Specialist, Wye Research & Education Center)
Design, Layout and Editing: Suzanne Klick (Technician, CMREC)

Damage From Box Tree Moth Caterpillars

By Laura Nixon

This week I scouted two sites in Western Maryland which had some box tree moth (*Cydalima perspectalis*) caterpillar activity last week. At my first site in **Hancock, MD**, there were only empty hibernacula (overwintering webbed structures) and actively feeding caterpillars; all the caterpillars at this site were close to 1” long and at the 5th and 6th larval instar. At the second site in **Clear Spring, MD**, about half of the hibernacula I opened still contained overwintering larvae. The caterpillars that were out and feeding varied from 0.4” – 0.8”, indicating a mix of 3rd to 5th instars. At both sites, fresh “curlicue” damage was already visible from the caterpillars defoliating the boxwood leaves. It is important to observe the life stage of the caterpillars present, as many treatments, such as Btk (*Bacillus thuringiensis* kurstaki), are more effective on the younger instars.



Box tree moth caterpillar active in Berkeley County, WV. This is likely a 4th or 5th instar larvae; box tree moth have 6 or 7 instars.

Photo: Austin Merrbaugh, Antietam Tree and Turf

Additional reports of box tree moth activity came in this week from

Dover (Kent County), DE and Berkeley and Jefferson Counties, WV. These counties are adjacent to Queen Anne's and Washington Counties, MD, respectively. For treatment recommendations, see the [March 5th Special IPM Alert](#).



“Curlicue” damage where box tree moth has fed on the boxwood leaf and left only the outer edge behind.

Photo: Laura Nixon, UME



Box tree moth caterpillar in Dover, DE. Note the webbing and frass surrounding this caterpillar, these are additional signs to look for when scouting.

Photo: Chris Griswold, Plant Health Care Specialist



Box tree moth at different larval stages taken from the same boxwood plant in Washington County, MD.

Photo: Laura Nixon, UME

Boxwood Spider Mite Hatch

By Laura Nixon

The erratic temperatures may have slowed down some of our early season pests, but boxwood spider mite (*Eurytetranychus buxi*) eggs hatch at 141 DD, which has been exceeded throughout Maryland this week. This mite overwinters as clear-colored eggs which turn red just prior to hatch, and will currently be in the larval or nymphal stages. You may be seeing heavy stippling on your boxwood leaves from last year, or the beginning of fresh yellow stippling from this year.

Now is a good time to treat those early life stages of the mite. Hexythiazox (Hexygon), a mite growth regulator, or 2-3% horticultural oil is effective at this timing before summer populations get going. If you have a mild infestation or treat later in the Spring, it is best to stick to a lower 1-2% oil rate. These treatments require complete plant coverage to be most effective, as they are contacts.



Boxwood mite damage on foliage.

Photo: Suzanne Klick, UME

Be Prepared for Boxwood Leafminer Adult Emergence

By Laura Nixon

Last week, we reported boxwood leafminer (*Monarthropalpus flavus*) larvae preparing to pupate. Many regions of Maryland have reached the 249 DD threshold for emergence of the adult leafminers this week. Adults are orange colored and resemble mosquitoes. If you observe high numbers of adults emerging over the next week, they can be treated with contact sprays. However, the window of time to effectively treat adult boxwood leafminer is very narrow, and treating the next generation of larvae is more effective. For details on treatment options, see the [April 3rd 2026 IPM report](#) and the [October 10th 2025](#) report.



Yellow to brown areas on boxwood foliage can indicate late instar and pupal activity of boxwood leafminer.

Photo: Suzanne Klick, UME



A boxwood leafminer adult on the underside of a boxwood leaf.

Photo: Joe Bogg, Ohio State University Extension.

Maskell Scale on Leyland

By Laura Nixon

Colin Campbell, Bartlett Tree Experts, observed a heavy scale infestation on Leyland cypress. Despite minute cypress scale and juniper scale being the more common species seen on Leylands, this scale sample appears to be Maskell scale (*Lepidosaphes pallida*). Maskell scale is an armored scale, which overwinters as mated females (pictured). These scale are oyster-shaped and light brown; if the armor is flipped the female body below is white. Crawlers emerge at 470 DD, which tends to be in June, and are pale yellow or white. There is a second generation of crawlers in Maryland in late summer.

Maskell scale are commonly found on cryptomeria, black pine, and junipers. When populations are high on a plant, the symptoms can appear as foliage die-back or needle browning depending on the plant species. Crawlers can be treated with horticultural oil, an insect growth regulator (buprofezin [Talus] is labelled for Maskell scale), or other labelled contact insecticides. Monitor the population after treatment, and, if it persists, systemics such as cyantraniliprole (Mainspring GNL) or dinotefuran (Safari, Venom) can be used. Be mindful not to apply systemic and contact insecticides with pollinator warnings around bloom time.



Maskell scale was found on Leyland cypress. Photo: Colin Campbell, Bartlett Tree Experts

Boxelder Bug Adults are Abundant and on the Move

By: Paula Shrewsbury

On Wednesday, April 8th, I was at a playground in Columbia, MD and there were hundreds to thousands of boxelder bug, *Boisea trivittatus* (Rhopalidae), adults running all over the place – on the swings and other play equipment, mulched ground, turf, trees, rocks, and more. Boxelder bugs have left their protected overwintering sites (ex. your homes, outdoor locations) and are now in search of mates and food. It is not unusual to find this level of box elder bug activity in areas where maples and boxelders are present – which they were at the Columbia playground. There were many mating pairs of bugs and females will soon lay eggs on the bark of trees or on the ground. Boxelder bug nymphs and adults feed on the developing seeds of boxelder and those of other maple species (Family Sapindaceae).

Boxelder bugs are a nuisance bug. They do not usually cause visible damage to trees and do not require control. Areas where you see adults flying and crawling now should be monitored in late summer to early fall for aggregations of boxelder bugs that may find their way into homes to overwinter (nuisance pest).



A mating pair of boxelder bugs recently emerged from their overwintering location.

Photo: Paula Shrewsbury, UMD



Boxelder bug adult on the bark of a maple tree.

Photo: Paula Shrewsbury, UMD

Roses Growing Strangely? It could be rose rosette disease

By Ana Cristina Fulladolsa

Rose rosette virus can infect almost all rose cultivars and species. Initially, stems may appear elongated and yellow to reddish. Many rose cultivars have a reddish coloration, particularly in young stems, but they will generally turn green as they continue to grow. In plants infected with rose rosette virus, the tissue remains reddish and appears malformed. Excessive thorns are commonly observed on stems and are flexible when they start to grow. Other symptoms include proliferation of small leaves and stems in tight groupings (witches-broom or rosette), yellow or red leaf mottle, succulent and thickened stems, decreased winter hardiness, and increased susceptibility to other diseases.

Roses should be regularly inspected for symptoms of rose rosette disease. If you observe symptoms, uproot and remove plants immediately to avoid virus spread to other plants. The virus is transmitted by tiny eriophyid mites that survive in infected material and move to other plants through wind, rain, tools, etc. Pruning symptomatic material is not effective because the virus spreads quickly to all plant parts, including roots.

To minimize the risk of rose rosette disease, plant other species between roses and space out rose bushes. Avoid using leaf blowers close to roses as they may house mite vectors that can easily be blown to nearby plants. If multiflora roses or other wild roses are found in the vicinity, remove them or place ornamental plantings upwind of the wild roses and avoid contact with their foliage.



Rose rosette disease symptoms on stems and leaves.
Photo: Jennifer Olson, Oklahoma State University, Bugwood.org.



Multiflora roses with rose rosette disease symptoms, found in a natural area in Montgomery County.
Photos: Karen Rane, 2026.

The Roseslug Sawfly – One and Done

By Laura Nixon

The roseslug sawfly (*Endelomyia aethiops*) is one of three common sawflies that attack roses (the others are curled and bristly rose sawflies). Unlike the other two species, the roseslug sawfly only has one generation per year. The early instar larvae are active after 240 DD, so be scouting your roses for signs of early damage in the next 1-2 weeks. These larvae scrape and chew the top layer of the leaf, causing windowpane damage, and only skeletonize leaves when populations are high. The larvae will feed into early summer, then drop into the soil to spin cocoons and prepare for overwintering. Damage from this species tends to be cosmetic rather than harmful to the plant.



Roseslug sawfly larva.

Photo: John A. Weidhass, Virginia Polytechnic Institute and State University, Bugwood.org

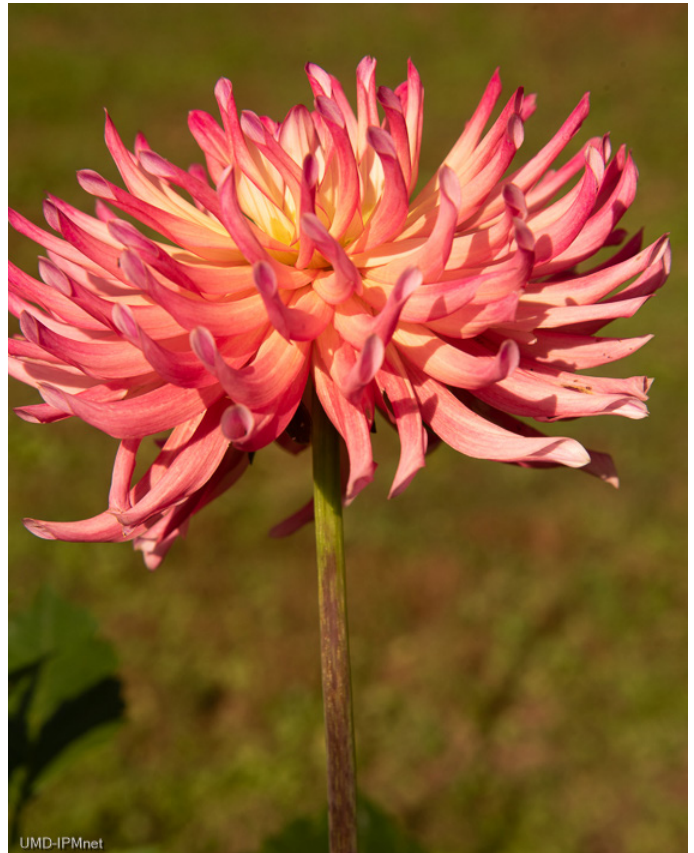
Prune out small sections of the rose which has active sawfly larvae or sawfly damage.

If populations persist and the damage is significant to the plant value, larvae can be treated with any contact or systemic labelled for sawflies. However, populations are generally kept down by natural enemies, so avoid spraying broad spectrums around your roses.

A New Era for Greenhouse and Cut Flower News!

Next Wednesday (15th April), we will begin sending out the Greenhouse and Cut Flower Newsletter again. We will start on a monthly basis at this time and send out alerts or conference announcements as necessary. The new version of this newsletter will be coordinated by Laura Nixon (Lnixon1@umd.edu, UME) and our colleague at PennState, Patricia Prade (prade@psu.edu), and cover updates from both Maryland and Pennsylvania.

If you were previously subscribed to the Greenhouse Newsletter, you are still subscribed! If you have not previously subscribed and have interest in greenhouse production and floriculture, you can subscribe on [our greenhouse/cut flower alert web page](#).



Find out what is showing up on dahlias in the first report.
Photo: Suzanne Klick, UME

Oh No... Not another insect on boxwood!

By: Paula Shrewsbury and Nancy Harding

Given all the pest insect and disease problems that occur on boxwoods, they have been in the UME IPM Reports a lot. Unfortunately, I have one more pest insect to add to the list, a somewhat new species of psyllid, *Spanioneura fonscolombii* (Hemiptera: Psyllidae). Sorry, no common name for this one yet. This psyllid is in addition to the boxwood psyllid, *Spanioneura buxi* (formerly *Psylla buxi*) that has been on our screen as a pest of boxwood for a long time. Both psyllids are non-native, which is not surprising since they specialize on boxwood (*Buxus* spp., Buxaceae) as their host plant and boxwoods are non-native to the U.S., but heavily integrated into managed landscapes.

Spanioneura buxi (formerly *Psylla buxi*) is commonly referred to as boxwood psyllid or box psyllid. It is thought to be native to Europe and was first detected in U.S. in 1884. It now occurs throughout U.S. wherever boxwoods occur. *Spanioneura fonscolombii* was first reported in Connecticut in 1916, but interestingly later surveys of psyllids in the U.S. state *S. fonscolombii* had not been detected since the initial U.S. 1916 discovery. As the story continues, in 2012 Nancy Harding (Entomology, UMD) re-discovered *S. fonscolombii*. Nancy brought samples from boxwoods (*Buxus sempervirens*) in her yard (Bowie, MD) to the Shrewsbury lab and stated she thought there was a different psyllid on her boxwoods (adults looked different and far more white wax on the branches and leaves). We brought a psyllid sample to scientists at USDA in Beltsville where the psyllid identification was confirmed to be *S. fonscolombii*, not *S. buxi*. Since then, there have been additional reported sightings of *S. fonscolombii* in Washington D.C. and other locations in Maryland (pers. obs.) and in online databases for community scientists in several other states, including Connecticut, Maryland, Massachusetts, New Hampshire, New Jersey, Pennsylvania, Rhode Island, and Virginia (iNaturalist community 2018-2025). *Spanioneura fonscolombii* and *S. buxi* may co-occur together on a boxwood. Though throughout the 15 month-long sampling period in a study we conducted on boxwoods, only three *S. buxi* were recorded, whereas 2,095 *S. fonscolombii* were recorded in total.



Images of psyllid life stages: 2) *Spanioneura fonscolombii* adult, note the diagnostic 4 dark spots on the edge of the forewing; 3) *S. buxi* adult; 4) *S. fonscolombii* nymphs and white wax; 5) Boxwood leaf bud scale showing eggs of *S. fonscolombii*;
Photos: Nancy Harding, UMD; from Russavage et al, subm. for publication

Similarities and differences between *S. buxi* and *S. fonscolombii*:

Adults are similar in size and color. The identifying characteristic is the presence of four distinct dark spots along the apical (front) margin of the front wings of *S. fonscolombii* (see image), which are not present in *S. buxi* ([go to iNaturalist for more images](#)). **Damage** by *S. buxi* displays as cupping distortion of the new leaves with white wax present on those leaves when nymphs are active; nymphs are most active on the new growth. In most cases but not all, this damage does not affect boxwood health and is not at a level where control measures are necessary. Damage by *S. fonscolombii* expresses as very high levels of white wax present on the leaves and stems of boxwoods, much more wax than I have ever seen from *S. buxi*; nymphs feed on the new growth and stems of boxwood. The good news is that, to date, other than heavy waxing, no direct feeding damage has been noted from *S. fonscolombii*. Both psyllids have one **generation** per year. *Spanioneura buxi* **overwinter** as orange-colored eggs in the buds of boxwood; while our research found that *S. fonscolombii* spend the winter as adults (likely somewhere in the plant or in leaf litter on the ground); and begin laying eggs in tight leaf buds of boxwood (~late February to early March). Nancy noted that *S. fonscolombii* **eggs hatched** around the end of March (DD ~120-132) in Bowie, MD. This week she observed lots of white wax and active 1st instar nymphs. Our research indicated nymphs were active until mid-May in MD. *Spanioneura buxi* eggs hatch ~ 184 DD (DD in MD this week ranged from 157 – 259 DD depending on location; see the table at the end of this report). Adults were active throughout the year.

We suspect that *S. fonscolombii* may have a broader range in the U.S., but many people “assume” what they have been seeing is boxwood psyllid (*S. buxi*). If you have boxwoods with heavy white wax being produced, this may be the “new” *S. fonscolombii* psyllid. If you think you have this one, please take a picture and email it to me (pshrewsbury@umd.edu) with the date and location. If you can get a sample or close up image of the adult psyllid, that would be great too!



Spanioneura fonscolombii adult on a fingertip for relative size collected from boxwood this past week in Winchester, VA.

Photo: Brian Ruether, USDA



Abundant wax produced from *Spanioneura fonscolombii* nymphs on boxwood.

Photo: Nancy Harding, UMD

Beneficial of the Week

By: Paula Shrewsbury

Zebra swallowtail butterflies – pollinators of Paw paw trees

Last Saturday, April 4th, was a warm day and I was riding my bike along the C&O Canal path in the Sharpsburg, MD area. I observed my first Zebra swallowtail butterfly, *Protographium marcellus* (Lepidoptera: Papilionidae), adult of the season. To my excitement, I further observed at least 15 more “Zebras” flying around that day. Zebra swallowtails are native to the eastern, central, and southern regions of the U.S. Zebra swallowtails are the only non-tropical species of kite-swallowtails (tribe of swallowtails they belong to) in its range.

Along the bike trail, I also observed that Paw paw trees, *Asimina triloba*, were in full bloom displaying their interesting, maroon and green-colored flowers. Paw paw is an understory tree native to Eastern North America and belongs to a plant family referred to as the tropical custard-apple family (Annonoaceae). Paw paw is the only plant in its family and genus in the temperate zone (all others are tropical), and it produces the largest edible tree fruit native to the U.S. Paw paw fruit are highly nutritious and can be eaten fresh, or included in ice cream, smoothies, baked goods and other foods. In the last 15 or so years, paw paws have grown considerably in popularity. What does this mean to our industry? More homeowners want to grow paw paw trees in their yards, and more nurseries are producing paw paws.

It was not a coincidence that Zebra swallowtails were active when Paw paw trees were in bloom. Zebra swallowtails and Paw paw have a close biological relationship. Zebra swallowtails are important pollinators of Paw paw, along with some beetle species. Paw paw flowers emit a slight carrion-like odor that attracts Zebra swallowtail butterflies. The butterflies feed on nectar and travel between flower of different Paw paw plants, aiding in cross-pollination (paw paws are self-incompatible). Zebra swallowtail butterflies also pollinate blueberry, blackberry, lilac, redbud, verbena, dogbane, and common milkweed. The other part of the Paw paw – Zebra swallowtail relationship involves the caterpillar stage of the Zebra swallowtail. Zebra swallowtail caterpillars are specialist herbivores and only eat Paw paw foliage, with a preference for young foliage. Female Zebra butterflies lay individual eggs near growing points where new leaves are expanding. Zebra swallowtails overwinter as pupae (chrysalis that look like a rolled leaf) on the underside of leaves or stems of Paw paw. In most of the Zebra swallowtail range, there are two generations per year. Densities of Zebra swallowtail caterpillars are not known to ever be high enough to cause heavy defoliation to Paw paw. To this point, for years whenever I come across Paw paws with



A Zebra swallowtail butterfly laying an egg on a newly expanding leaf of a Paw paw tree. Note how the abdomen is curled forward with its tip on the leaf.

Photo: BudOhio, CC BY-NC-ND 2.0 vis Flickr.com



Partially open flower bud of Paw paw taken on April 4, 2026.

Photo: P.M. Shrewsbury, UMD

signs of defoliation, I stop to search the foliage in hopes of finding a Zebra swallowtail caterpillar. Much to my dismay, I have not found a caterpillar yet!

Zebra swallowtails are beautiful native butterflies that pollinate a native edible fruit. Both the butterfly and the plant are unique among their taxonomic groups, in that they are the only ones that grow in temperate regions (all others are tropical species). I would say Zebra swallowtails are very beneficial insects. [Click here to see a YouTube video of Zebra swallowtail adults puddling to get sodium and minerals, and caterpillars \(by M.J. Raupp, Bug of the Week\).](#)



Flowering paw paws in Centreville, VA this week.
Photo: Dave Freeman, Oaktree Property Care



Zebra swallowtail caterpillar on Paw paw.
Photo: Seig&Alice Kopinitz, CC BY-NC-ND 2.0 vis Flickr.com



A Zebra swallowtail chrysalis (rolled leaf mimic) attached to the underside of a Paw paw leaf.
Photo: Judy Gallagher, cc-by 2.0 via flickr.com

Weed of the Week

By: Kelly Nichols

Large crabgrass seedlings are emerging here in central Montgomery County, MD. Large crabgrass (*Digitaria sanguinalis*) is a summer annual weed very common in lawns, landscapes, fields, and roadsides. The first leaf to appear will be wide and short in length (Figure 1). As the seedlings grow, the membranous ligule and hairs on the leaves and stems (Figure 2) will become more evident. The seedhead is finger-like, hence the “digit” in “Digitaria” (Figure 3). Smooth crabgrass (*Digitaria ischaemum*) looks similar but has no hairs and is an overall smaller plant.

Post-emergent products for crabgrass control include quinclorac (Drive), mesotrione (Tenacity), and combinations of quinclorac plus carfentrazone (SquareOne) or sulfentrazone (Solitaire). For non-synthetic options, chelated iron (Fiesta) provides some control; higher rates applied closer together will likely be needed. In a 2024 study at the University of Maryland's Turfgrass Research Center in College Park, the high rate of the granular formulation (8 lbs/1,000 sq ft) provided 80-85% control when applied twice at 3-, 4-, or 6-week intervals. The mid-rate of the liquid formulation (12.6 fl oz/1,000 sq ft) provided 87% control when applied at a 4-week interval. For this study, the first application was on April 10, and the ratings mentioned here were taken 11 weeks later (end of June). It is also important to note that nitrogen fertilizer applied at a 4-week interval provided 88% control, indicating that a good soil fertility program and healthy turf are key for combating weeds like crabgrass.

Other components of a crabgrass management plan that should be prioritized are proper mowing height (3-4 inches) and maintaining optimum pH. Build a strong turf that is dense. This prevents sunlight from reaching the soil to allow germination of crabgrass seeds.



Figure 1. Large crabgrass seedling leaves are wide and short.
Photo: Rebekah D. Wallace, University of Georgia, Bugwood.org.



Figure 2. Large crabgrass stem and leaf with hairs.
Photo: Joseph M. DiTomaso, University of California - Davis, Bugwood.org.



Figure 3. Flowering finger-like seedhead.
Photo: Joseph M. DiTomaso, University of California - Davis, Bugwood.org.

Plant of the Week

By: Ginny Rosenkranz

Mertensia virginica or Virginia bluebells is one of the loveliest of the native spring ephemerals. The plants emerge from winter dormancy with deep purple foliage that quickly turns green in order to have the chlorophyll in the leaves absorb light energy from the sun to grow and produce flowers, and seeds to continue their species. The leaves are oval to oblong with prominent veins and entire or smooth margins. Leaves are longer at the base and grow smaller as they climb the arching stems in an alternate fashion. Most of us spend more time enjoying the arching stems with the flowers clustered on the ends. The flower buds start out with a soft pink color that changes to bright blue as the lightly fragrant flowers grow to 1-inch-long, bell-shaped blooms. Each flower becomes a nodding trumpet-shaped bloom by having the 5 petals fused together. Inside of each flower are 5 white stamens with light brown anthers for pollen. They surround the long white pistil in the middle of the flowers. Like most of the spring ephemerals, Virginia bluebells thrive under the shade of deciduous trees, catching the sunlight in spring before the leaves emerge. This allows the plants to bloom for up to 3 -4 weeks in the coolness of spring. While in bloom, the flowers provide nectar that is needed by many pollinators like bees, including the bumble bees and the long-tongued bees. Butterflies, skippers, moths, and hummingbirds also visit and pollinate the flowers, adding color and action in the spring gardens. Once finished blooming, the flowers again change color back to light pink, looking like deflated balloons. The 4-lobed fruits are called schizocarps and contain 4 nutlets per flower and help the plants expand into colonies. There are also Virginia bluebells that bloom with white or light pink flowers. The foliage dies around mid-summer, going dormant until next spring. While they are dormant, their space can be filled with other shade-loving perennials to enjoy until autumn-like native ferns and coral bells. Virginia bluebells are cold tolerant in USDA zones from 3-8, thriving in part to full shade. They prefer to grow in rich, moist soils, have no serious insect or disease problems and they are tolerant of rabbits and black walnut trees.



Blue flowering Virginia bluebells are more often seen, but there can be white flowers as well.
Photos: Ginny Rosenkranz, UME

Pest Predictive Calendar “Predictions”

By: Nancy Harding and Paula Shrewsbury

In the Maryland area, the accumulated growing degree days (**DD**) this week range from about **157 DD** (Clarksville) to **259 DD** (Nat'l Arboretum/Reagan Nat'l). The [Pest Predictive Calendar](#) tells us when susceptible stages of pest insects are active based on their DD. Therefore, this week you should be monitoring for the following pests. The estimated start degree days of the targeted life stage are in parentheses.

Boxwood spider mite – egg hatch (**141 DD**)
European pine sawfly – larva, early instar (**154 DD**)
Woolly elm aphid – egg hatch (**163 DD**)
Inkberry holly leafminer – adult emergence (**165 DD**)
Spiny witchhazel gall aphid – adult/nymph (**171 DD**)
Spruce spider mite – egg hatch (**179 DD**)
Boxwood psyllid – egg hatch (**184 DD**)
Tea scale – egg hatch / crawler (1st gen) (**195 DD**)
Hemlock woolly adelgid – egg hatch (1st gen) (**197 DD**)
Viburnum leaf beetle – first egg hatch (**210 DD**)
Azalea lace bug – egg hatch (1st gen) (**214 DD**)
Birch leafminer – adult emergence (**219 DD**)
Roseslug sawfly – larva/early instar (**230 DD**)
Elongate hemlock sciae – egg hatch / crawler (1st gen) (**232 DD**)
Boxwood leafminer – adult emergence (**249 DD**)
Hawthorn lace bug – first adult activity (**259 DD**)
Spotted lanternfly – egg hatch (**270 DD**)
Bristly roseslug sawfly – larva, early instar (**284 DD**)
Imported willow leaf beetle – adult emergence (**290 DD**)
Hawthorn leafminer – adult emergence (**292 DD**)
Andromeda lace bug – egg hatch (**305 DD**)
Pine needle scale – egg hatch / crawler (1st gen) (**307 DD**)
Cooley spruce gall adelgid – egg hatch (**308 DD**)
Eastern spruce gall adelgid – egg hatch (**308 DD**)

See the [Pest Predictive Calendar](#) for more information on DD and plant phenological indicators (PPI) to help you better monitor and manage these pests.

Degree Days (as of April 8, 2026)

Annapolis Naval Academy (KNAK)	163
Baltimore, MD (KBWI)	204
Belcamp (FS836)	162
Clarksville (001MD)	157
College Park (KCGS)	240
Dulles Airport (KIAD)	241
Ft. Belvoir, VA (KDA)	241
Frederick (KFDK)	180
Gaithersburg (KGAI)	210
Greater Cumberland Reg (KCBE)	200
Martinsburg, WV (KMRB)	211
Millersville (MD026)	219
Natl Arboretum/Reagan Natl (KDCA)	259

Perry Hall (C0608)	164
Salisbury/Ocean City (KSBY)	249
St. Mary's City (Patuxent NRB KNHK)	247
Westminster (KDMW)	248

Important Note: We are using the [Online Phenology and Degree-Day Models](#) site. Use the following information to calculate GDD for your site: Select your location from the map Model Category: All models Select Degree-day calculator Thresholds in: Fahrenheit °F Lower: 50 Upper: 95 Calculation type: simple average/growing dds Start: Jan 1

Conferences

May 20, 2026

Mid-Atlantic Nursery Workshop: the Water Loop to Pot: Managing Ponds, Irrigation, Substrates, and Runoff for Better Production Profits

Location: Hampton Roads Agricultural Research and Education Center, Virginia Tech, Virginia Beach, VA

[For more information on this program](#)

June 16, 2026

2026 Eastern Shore Procrastinators Conference

Location: Zoom

June 18, 2026

MNLGA Field Day

Location: Mt Cuba Center, Hockessin, DE

June 26, 2026

Montgomery County Pesticide Procrastinators Conference

Location: Derwood, MD

IPM Scouts' Diagnostic Sessions (1 - 3 p.m.)

June 23, 2026

July 22, 2026

August 26, 2026

Location: CMREC, Ellicott City, MD

Commercial Ornamental IPM Information

<http://extension.umd.edu/ipm>

CONTRIBUTORS:



Paula Shrewsbury
Extension Specialist
pshrewsb@umd.edu



Laura Nixon
Extension Specialist
lnixon1@umd.edu



David Clement
Plant Pathologist
clement@umd.edu



Ana Cristina Fulladolsa
Plant Pathologist
acfulla@umd.edu



Nathan Glenn
Extension Educator
Howard County
nglenn@umd.edu



Nancy Harding
Faculty Research
Assistant



Kelly Nichols
Extension Educator
Montgomery County
kellyn@umd.edu



Dan Buonaiuto
Assistant Professor
Dept. of Plant
Sciences and Land-
scape Architecture
dbuona@umd.edu



Andrew Ristvey
Extension Specialist
aristvey@umd.edu



Ginny Rosenkranz
Extension Educator
Wicomico,
Worcester, Somerset
Counties
rosnkranz@umd.edu

Thank you to the Maryland Arborist Association, the Maryland Nursery, Landscape, and Greenhouse Association, Professional Grounds Management Society, FALCAN, and USDA NIFA EIP Award # 2024700043556 for their financial support in making these weekly reports possible.

The information given herein is supplied with the understanding that no discrimination is intended and no endorsement by University of Maryland Extension is implied.

University programs, activities, and facilities are available to all without regard to race, color, sex, gender identity or expression, sexual orientation, marital status, age, national origin, political affiliation, physical or mental disability, religion, protected veteran status, genetic information, personal appearance, or any other legally protected class.