UNIVERSITY OF MARYLAND EXTENSION

TPM/IPM Weekly Report
for Arborists, Landscape Managers & Nursery Managers

Commercial Horticulture **April 3, 2026**

In This Issue...

Integrated Pest Management for Commercial Horticulture
extension.umd.edu/ipm

If you work for a commercial horticultural business in the area, you can report insect, disease, weed or cultural plant problems **(include location and insect stage)** found in the landscape or nursery to sklick@umd.edu

Coordinator Weekly IPM Report:
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Disease Information: David Clement (Extension Specialist) and Ana Cristina Fulladolsa (Plant Pathologist and Director, UMD Diagnostic Lab)
Weed of the Week: Kelly Nichols and Nathan Glenn, (UME Extension Educators) and Dan Buonaiuto, (Assistant Professor), Dept. of Plant Sciences and Architecture
Cultural Information: Ginny Rosenkranz (Extension Educator, Wicomico/Worcester/Somerset Counties)
Fertility Management: Andrew Ristvey (Extension Specialist, Wye Research & Education Center)
Design, Layout and Editing: Suzanne Klick (Technician, CMREC)

Box Tree Moth Caterpillars Are Active

By: Laura Nixon

Active box tree moth (*Cydalima perspectalis*) caterpillars have been found in Western Maryland this week. We scouted in locations which had populations in 2025; many people told us they pulled out boxwoods at the end of last year which had been completely defoliated and appeared dead. The box tree moth populations we found had some active caterpillars which had begun feeding on leaves and some caterpillars were still in their overwintering structures (called hibernacula). Spring emergence from overwintering hibernacula appears to be asynchronous and the caterpillars we found were of varying larval instars (~2nd-4th instar). As such, this Spring population is best treated with systemic materials with a long residual such as chlorantraniliprole (Acelepryn, Contrado SC) or cyantraniliprole (Mainspring GNL). Chlorantraniliprole is appropriate for bloom time.

See the previous 2026 alerts for box tree moth monitoring and treatment options. If you find box tree moth, report to UME (lnixon1@umd.edu, sklick@umd.edu) and MDA.

Boxwood with visible damage, some is last Fall's damage, but on closer inspection there is fresh feeding damage and frass.
Photo: Suzanne Klick, UME

Hibernaculum peeled open to reveal the overwintering larva (~2nd instar) that hasn't yet emerged.
Photo: Suzanne Klick

Box tree moth caterpillar (~4th instar) surrounded by webbing and frass.
Photo: Suzanne Klick, UME

Boxwood Leafminer

By: Laura Nixon

Marie Rojas (IPM Scout) reported boxwood leafminer (*Monarthropalpus flavus*) larvae beginning to turn orange this week, indicating that the insect is preparing to pupate. There are no chemical control measures necessary at this stage as the larvae have already done their feeding damage, most of which occurred last Fall. However, if you have a particularly heavily infested section on your plant, you can prune out those branches to reduce the number of adults emerging this Spring. Keep an eye out for pupation and adults emerging over the next 2 – 3 weeks. Once adults emerge, they can be treated with a contact spray, but that treatment window is very narrow. Data shows that treating with a systemic, such as imidacloprid (basal soil drench or foliar spray) or abermectin (foliar spray), once adults have oviposited is highly effective to target the next generation of larvae. Timing is important with these systemics, they should not be applied during or prior to bloom to protect pollinators. However, boxwood leafminer larvae go into aestivation during the summer and are not feeding, so mid to late Spring applications will be most effective.

**Peel open boxwood leaves with visible leafminer damage to see the larvae.
Photo: Marie Rojas, IPM Scout.**

**Two boxwood leafminer pupae extracted from a boxwood leaf.
Photo: Suzanne Klick, UME**

Eastern Tent Caterpillars

Eastern tent caterpillars continue to be active. Dave Lantz found tents in Boonsboro, MD on April 2. The best management method at this point is to use a stick to tear open the tents and expose the caterpillars to predators.

**Look for the tents of eastern tent caterpillars in the crotches of various trees.
Photo: Dave Lantz**

Scale on Aucuba

By Laura Nixon, UME

This week, Paul Wolfe (Integrated Plant Care) sent me an envelope full of scale! The sample was two cuttings from an Aucuba plant he manages which was covered in an armored scale. This scale is either oleander scale (*Aspidiotus nerii*) or false oleander scale (*Pseudaulacaspis cockerelli*), it can be hard to differentiate. Both are pale, circular armored scale; when flipped a darker yellow or brown body is beneath. Oleander scale is present in Maryland, but is not commonly a pest issue. False oleander scale is seen a little more commonly, especially on magnolias, and damage symptoms look like yellow, chlorotic spots on leaves from heavy feeding. For both species, start by pruning out heavily infested branches. If the pest persists and you begin to see plant damage, monitor for crawler activity and treat that stage with oil followed by a systemic if needed.

Oleander scale or false oleander scale is active on aucuba this week. The probing needle in the photo is pointing to where a female cover was flipped to show the body underneath.
Photo: Laura Nixon, UME

Cottony Camellia/Taxus Scale

Bill Miller, The Azalea Works, and Sam Fisher, Bartlett Tree Experts, found second instar overwintering females of cottony camellia/Taxus scale in Bethesda and D.C. this week. Both of them found this scale infesting camellias. Monitor scale infestations for crawlers around 649 degree day (usually late May to early June). Treat the crawlers at that time.

Cottony camellia/Taxus scale females darken as they mature.
Photo: Sam Fisher, Bartlett Tree Experts

A Perfect Storm: Part I

By: Andrew Ristvey

Last year, I visited a new grower who was experiencing significant plant health issues. The operation was a small container nursery using well water, producing a variety of woody plants in a very loose, porous pine bark substrate. The grower had been consulting with a well-known horticultural researcher and author, but despite this guidance, plants had begun to show both root decline and dieback in the canopy. Based on advice from the grower's local acquaintances who figured the root dieback was from overwatering, the grower reduced irrigation. However, conditions did not improve. In fact, the symptoms became more widespread, affecting an increasing number of plants.

I was invited to assess the situation. After a thorough walkthrough of the nursery, I began my usual diagnostic process. Given the symptoms, I first focused on the root systems, removing several plants from their containers for inspection. Root growth was noticeably limited compared to what would be expected since transplanting, but there were no clear visual signs of disease.

My next step was to conduct a saturated media extract (SME). While the test was processing, I began asking standard questions: What is your fertilizer program? How is irrigation managed? Where does your substrate come from, and how is it handled? I also asked about water quality, particularly alkalinity. As the grower responded, it became clear that multiple interacting issues were contributing to the problem.

Returning to the primary concern—poor root development and shoot dieback—the SME results were revealed that the substrate pH was approximately 3.8, which is excessively low, while the electrical conductivity (EC) was around 4.0, indicating a high level of soluble salts. Several factors had converged to create this situation: the use of slow-release fertilizer, daytime temperatures in the 90s, and reduced irrigation. Together, these conditions promoted salt accumulation in the root zone.

The substrate itself—primarily pine bark—was highly porous. Based on similar materials I've evaluated, I estimated the air-filled porosity to be around 35% by volume, which corresponds to relatively low water-holding capacity. Under these conditions, reducing irrigation was counterproductive. My recommendation was to increase irrigation frequency, applying shorter, more frequent cycles to both meet plant water demand and leach accumulated salts from the substrate. In this case, nutrient leaching was not a concern—it was necessary.

This situation highlights an important principle: irrigation management must align with substrate physical properties, particularly porosity. Growers use a wide range of substrate components—peat, pine bark, coir, wood fiber—each with distinct water-holding characteristics. The specific mix is less important than how irrigation is managed in response to those properties.

In general, the most effective irrigation strategy is to replenish water as it is lost through plant transpiration. In highly porous substrates, this often requires frequent, short-duration irrigation events to minimize leaching. In contrast, substrates with greater water-holding capacity can be irrigated less frequently and with longer application times.

In all cases, consider nutrient management alongside irrigation. While intentional leaching can be used to reduce excessive salt buildup, routine practices should aim to keep the leaching fraction—the proportion of applied water that drains from the container—below 15%. There are additional nuances to determining when and how much to irrigate, but those details are beyond the scope of this discussion. For those interested in a deeper dive, I have co-authored an article with Dr. Gerry Spinelli on irrigation timing that will appear in Greenhouse Management this month. Keep an eye out for it. I'll be back later this month to discuss more of the issues in A Perfect Storm!

Cedar-apple Rust Galls Are Releasing Spores!

By: Ana Cristina Fulladolsa and David Clement

With recent rains and spring temperatures, we will start to see cedar-apple rust (*Gymnosporangium juniperi-virginianae*) sporulating on Eastern red cedar and junipers (*Juniperus* spp.). Infected cedars and junipers will develop greenish-brown galls that produce orange, gelatinous, tentacle-like structures that can enlarge to be several times longer than the size of the gall. These structures are called telial horns and they release spores that travel by wind and can infect susceptible apple and crabapple trees nearby.

Telial horns of cedar-apple rust produced on a juniper gall. Photo: Dave Clement, UMD

Cedar-apple rust requires two hosts to complete its life cycle: a cedar/juniper host and an apple/crabapple host. Generally, the fungus does not cause significant damage on the cedar/juniper host, but it can be devasting on the other host. Once spores released from the cedar/juniper host infect susceptible apple/crabapple, symptoms will develop on the latter, including yellow to bright orange or red leaf spots, premature defoliation, and production of smaller or malformed fruits. Fruiting structures of the rust fungus may form and appear as black spots within leaf spots on the upper surface of the leaves. On lower leaf surfaces, fringe-like projections may form and release spores that can then infect the cedar/juniper host.

The most effective way to manage this disease is to separate the hosts. This is usually impractical because ideal separation between hosts is 1-2 miles and there may be cedar/juniper plants in surrounding areas. If you are planning to install new trees in a landscape, avoid planting both hosts in the same are or use a resistant apple/crabapple cultivar. If you see telial horns on galls of cedar or juniper, protect susceptible apple/crabapples with preventive fungicides. This must be done before leaf spots are observed. If symptoms occur on the apple/crabapple host, prune infected twigs and branches and remove them from the area.

Fungicides are not effective in controlling infections on the cedar/juniper hosts. The fungus overwinters in galls, which dry up in late fall-winter. At that time, prune branches at 4-6 inches from the galls and remove all infected material from the area.

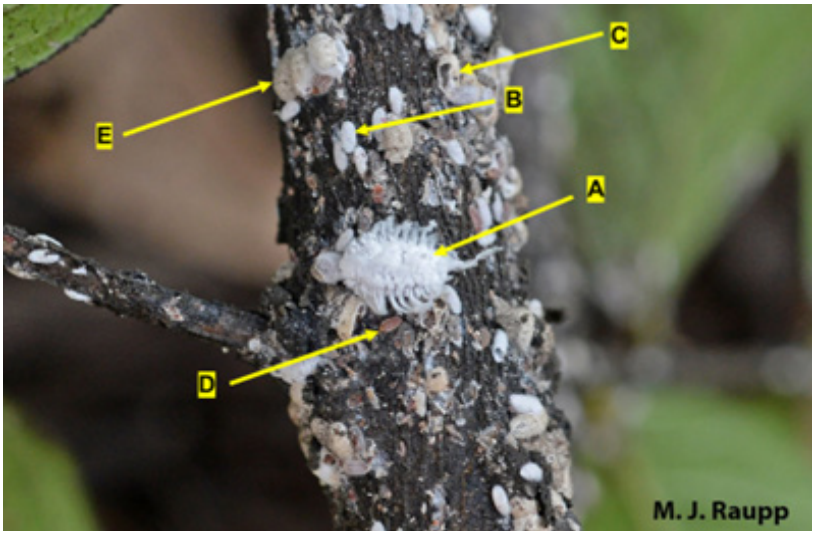
You can learn more about cedar-apple rust and its management at the websites listed below:

UMD Home and Garden Information Center: https://extension.umd.edu/resource/rust-diseases-trees/
Penn State Extension: https://extension.psu.edu/cedar-apple-and-related-rusts-on-ornamentals

Crapemyrtle Bark Scale is Active

By: Paula Shrewsbury, UMD

Sheena O'Donnell (UME Research Tech., CMREC) is continuing to monitor the life stages and densities of crapemyrtle bark scale (CMBS) so we can have two years of data (2025, 2026) on the number of generations and crawler activity times of CMBS in MD. Monitoring crape myrtles on April 1st in College Park MD, Sheena found that overwintering stages are now active and continuing to feed and develop. She saw mostly 2nd instar scales, but also some adult females and male pupal cases. No eggs or crawlers yet. We will continue to monitor weekly and let you know when crawler activity begins to occur and peak. Last year the first peak in crawler activity occurred around 966 DDs.



Crapemyrtle bark scale life stages and a *Hyperaspis* lady beetle larva. *Hyperaspis* lady beetle larva (A) feeding on crapemyrtle bark scale (CMBS); CMBS male pupal case (B); CMBS female ovisac that has been fed on (C); CMBS crawler (D); and CMBS intact ovisac made by female CMBS (E).
Photo: Mike Raupp and Paula Shrewsbury, UMD

Recommendations: Given the above observations, I recommend you monitor crape myrtles with CMBS and confirm they are alive or dead, and the life stages on the trees. Be sure to use a hand-lens or other magnification. Live CMBS will be pink to purple in color and will gush this color when you squish them, dead ones won't gush. Depending on the density of the population and how many trees you are managing, you can use mechanical control such as a soft scrub brush and water to physically wash the scales off the branches and trunk of the tree which can help to significantly reduce the populations. There are also multiple chemical controls that are available for CMBS suppression that should target the crawler stage for best results. These include systemics such as dinotefuran, insect growth regulators such as pyriproxyfen or buprofezin, or contact products such as horticultural oil or other labeled products. Be sure to follow label directions to protect pollinators and get optimal control. I found a very good fact sheet on CMBS from Clemson Cooperative Extension that anyone who is dealing with CMBS might want to read. It provides a thorough description of CMBS biology and management.

White Pine Weevil Adult Activity – Time to protect the tree terminals

By: Paula Shrewsbury

On April 1st this week in Frederick MD, Marie Rojas (IPM Scout) found oozing sap (white resin) on the terminals of *Picea omorika*, indicating feeding activity of adult white pine weevil, *Pissodes strobi*, has started. White pine weevils overwinter as adults in the leaf litter under host trees. First adult activity is predicted to be around 84 DD – which most areas in MD have reached and/or passed this week (see DD accumulations at the end of this report). White pine weevil is primarily a pest of eastern white pine, Colorado blue, Norway, and Serbian spruces. Scots, red, pitch, jack, and Austrian pines, and occasionally Douglas-fir are also attacked. This is a pest in nurseries, landscapes, and X-mas tree farms. Adult white pine weevils overwinter in litter on the ground or in old pine tree stumps. As weather warms, the adults begin feeding on terminal growth of conifers causing sap to ooze. They tend to feed on branch terminals about 10" below terminal buds. Females chew small holes in the bark of the terminal growth and deposit eggs in the holes causing the sap to flow (white pitch), and eggs hatch in 7-10 days. The developing larvae bore down within the terminal branches until they reach maturity around the end of June. Ultimately, the terminals take on the characteristic "Shepard hook" appearance and die. The white pine weevil often kills 2-3 years of terminal growth.

Recommendations: If trees are showing signs of damage and/or trees had problems with white pine weevil last year, consider protecting the terminal branches with an insecticide application. NOW is the time to treat before adults can lay eggs in the wood. For control, products that contain bifenthrin or permethrin can be applied. Indoxacarb (Avaunt) insecticide is labelled for weevil control in nurseries. Another tactic is to prune out and destroy flagged terminals in June before the adults emerge.

Heavy sap flow (resin, pitch) on the terminal of spruce from white pine weevil adult feeding. Photo: Marie Rojas, IPM Scout

Ambrosia Beetle Update

Marie Rojas, IPM Scout, is reporting that ambrosia beetles were found today, April 3, boring into yellowwood trees in Montgomery County. In our trap here at the research center in Ellicott City, MD and from one in Chestertown, MD, we are finding *Xyleborinus saxesenii* and at least one other species that still needs to be identified. Temperatures are fluctuating a lot this spring. Monitor tree trunks closely for wet areas indicating ambrosia beetles are entering the tree. When ambrosia beetles are active, permethrin (e.g. Astro, Permethrin Pro) and bifenthrin (eg. Onyx) are registered for use on tree trunks in the landscape. For field-grown trees in nurseries, permethrin (e.g. Perm-up) is an option. Bifentrhin (OnyxPro) is labeled for use to tree trunks in landscape and nursery sites.

Beneficial of the Week

By: Paula Shrewsbury

Firefly larvae climb trees?

This week Caitlin Beckjord (Howard County Rec. and Parks) sent in photos of firefly (a.k.a. lightning bugs) larvae that she found tucked in between the bark ridges of a hackberry tree in Columbia, MD. Great find Caitlin! There are approximately 30 species of fireflies (Lampyridae) reported in MD. I am not sure which species of firefly larvae were on the hackberry.

Fireflies are beetles in a group known as soft-winged beetles in the order Coleoptera and the family Lampyridae. Most light flashing firefly species occur east of the Mississippi River, are active at dusk into the evening. Adults and larvae of many firefly species exhibit bioluminescent – they glow in the dark! The purpose of this bioluminescence varies. It is believed that flashes by adult fireflies are part of a signaling

system for attracting mates. Both male and female adults emit light intermittently or in specific flash patterns. However, it is not just adult fireflies that exhibit bioluminescent (glow in the dark), the larvae also glow. The common eastern firefly, Photinus pyralis, eggs are laid in moist soil and hatch about a month later. Immature fireflies are called glowworms (see image). They also emit light, though it is more of a low intensity glow that is believed to be used as a warning signal telling predators not to eat them as they are mildly toxic and taste nasty. Larvae of our eastern firefly develop over two summers, so they overwinter twice before pupating and emerging as adults sometime in May or June. Larvae of fireflies live and feed in the soil. Depending on species, fireflies may pupate in the soils (preferably rich in organic matter), leaf litter, rotting logs or the furrows in the bark of tree trunks. Hence, the larvae that Caitlin found hunkered down in the bark furrows of hackberry likely were getting ready to pupate.

An adult firefly (*Photinus* sp).
Photo by: David Cappaert, Bugwood.org

Why are fireflies considered beneficials? The soil active glow worms are voracious predators of soft-bodied invertebrates and known to feed on slugs, snails, worms, and other soil-dwelling insects. Glow worms use their mandibles to inject prey with a paralyzing neurotoxin, making it defenseless, and then secrete digestive enzymes that liquify the prey making it easier to consume. Click here to see a great YouTube of glow worms in action (by M.J. Raupp, UMD).

This would be a good time to search the bark of trees for larvae (or maybe pupae) of fireflies. I know I am going to look for them!

A glowworm, larva of a firefly, on the bark of a hackberry tree.
Photo: Caitlin Beckjord, Howard County Rec. and Parks

A firefly larva (glowworm) removed from a tree trunk to show the relative size.
Photo: Caitlin Beckjord, Howard County Rec. and Parks

Weed of the Week

By: Dr. Dan Buonaiuto

As our weather seems to be bouncing daily between winter-like gloom and summer-like bounty. You may be noticing carpets of green sprawled across forest floors and woodland transition areas. What you are likely seeing is lesser celandine, *Fica verna*, a rapidly spreading invasive plant in the buttercup (Ranunculaceae) family. Lesser celandine used to be a popular groundcover that was prohibited from sale in Maryland in 2017, but continues to spread in the state through natural dispersal.

Figure 1: Lesser celandine covering a wooded area. Photo: D. Buonaiuto UMD

Lesser Celandine Identification:

- Growth habit: A low growing (up to 4") perennial that forms dense mats (Figure 1).
- Leaves: Simple, glossy, kidney to heart shaped or arrow shaped with short stalks and irregular wavy edges.
- Flowers: Glossy buttery yellow flowers with 8-12 petals on stalks that extend above the leaves (up to 9", Figure 2).
- Seeds/fruit: The seed is a fuzzy achene, but this plant also reproduces through tiny pale bulbets that grow on the stem and underground tubers.
- Look-alikes: The native marsh marigold (*Caltha palustris*). The leaf margins of marsh marigold are finely toothed, with they are smooth on lesser celandine.

Figure 2: Lesser celandine flower Photo: D. Buonaiuto, UMD

Lesser Celandine Biology/Ecology:

This perennial plant is a spring ephemeral, emerging early in the spring before tree canopies fill in. The above ground portion dies back by early summer. Lesser celandine can reproduce through seed (though this is rare), bulblets (small aerial tubers on the stem that are dispersed when the above ground portion of the plant dies back) or through underground tubers. This plant is often associated with floodplains and woodlands, but can occur in turf and gardens (Figure 3). Flowering typically occurs in our region from March to May.

Lesser Celandine Management:

Mechanical control:

Mechanical control is difficult. Small patches can be dug up as long as you make sure to get the entire plant, including the underground tubers which can resprout, and stem-bourne bulblets which can be scattered during removal. Plant materials should be bagged in plastic before disposal to prevent secondary spread.

Chemical control:
Glyphosate can be used early in the season as long as temperatures are above 50° and there is no precipitation in the forecast for 12 hours. Other chemical options include products that contain at least two effective herbicides including triclopyr, 2,4-D, dicamba and MCPA. The seasonal window for chemical control is short—from when plants start flowering until about 50% of plants are flowering, usually by early April. Waiting beyond that can damage emerging desirable plants. Care should be taken when using these products near ornamentals as they can volatilize and cause damage. Sulfentrazone products, usually used for control of nutsedges have been reported to be effective on lesser celandine as well.

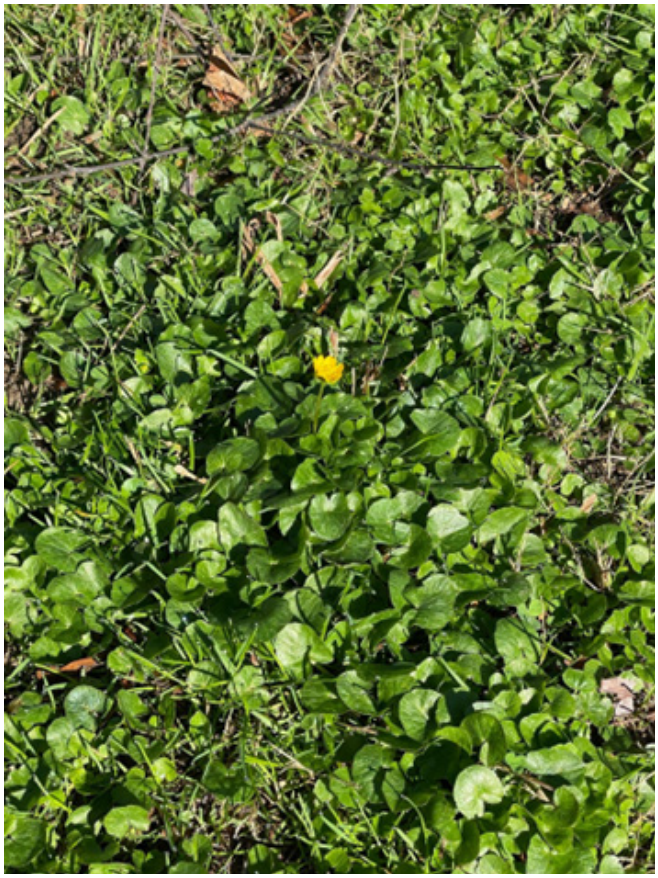
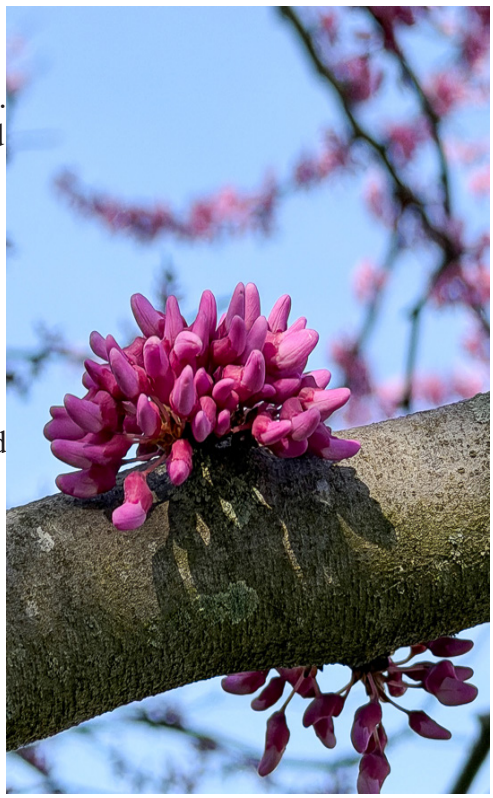


Figure 3: Lesser celandine can also occur in turf and garden areas.
Photo: D. Buonaiuto, UMD

Plant of the Week

By: Ginny Rosenkranz

Cercis canadensis or Eastern redbud is a native tree that can grow 20-30 feet tall and 25-35 feet wide. It prefers to grow in full sun to afternoon shade and loves to live in fertile soils with regular and consistent moisture. It doesn't like to have their roots disturbed, so the plants should be planted in their permanent location and kept well-watered until established. The plants can grow with a single trunk or can also grow into a multi-trunked tree. Most of the Eastern redbuds bloom a long time in early spring, from March through May before the foliage appears. The flowers grow in clusters directly on the branches of the trees, blooming on new growth as well as mature growth. The showy, pea-like flowers are only ½ inch wide with about 7 flowers to a clutter and come in light purple to lavender to rose pink with red stems and attract many pollinators. The brightly colored flowers are edible and can taste like peas, containing high amounts of vitamin C. The flowers mature into a flattened, bean-like, dry seedpods that grow 2-4 inches long with 6-12 seeds per pod. Once the blooms are done, the trees' heart-shaped, dark, lustrous green leaves appear in an alternate fashion on the dark reddish brown to black stems. The leaves have an entire margin and grow 3-6 inches long and wide and 7 palmate



Close-up of *Cercis canadensis* buds.
Photo: Ginny Rosenkranz, UME

veins. Autumn brings an attractive yellow fall color. There are 12 species of Lepidoptera that use the Eastern redbud as their larval host plant like Henry's Elfin butterfly, as do leafcutter bees (*Megachille rotundata*). butterflies including Henry's Elfin butterfly, bees, hummingbirds, and other pollinators feed on the nectar of the bright flowers. Native song birds and some small mammals feed on the seeds in the winter. These lovely native trees are cold hardy from USDA zones 4-8 and are tolerant of deer, clay soil, and black walnuts. There are several diseases that can harm this native tree such as Verticillium wilt, dieback, leaf spots, mildew, and blights. Insect pests can include Japanese beetles, treehoppers, leafhoppers, caterpillars, borers, webworms, and scale. Good soil moisture, light fertilization and proper pruning of dead branches help keep these beautiful trees healthy.

Overall growth habit of a redbud on a golf course. Photo: Ginny Rosenkranz, UME

Pest Predictive Calendar "Predictions"

By: Nancy Harding and Paula Shrewsbury

In the Maryland area, the accumulated growing degree days (DD) this week range from about 113 DD (Annapolis) to 197 DD (Nat'l Arboretum/Reagan Nat'l). The Pest Predictive Calendar tells us when susceptible stages of pest insects are active based on their DD. Therefore, this week you should be monitoring for the following pests. The estimated start degree days of the targeted life stage are in parentheses.

White pine weevil – adult first activity (84 DD)
Eastern tent caterpillar – egg hatch (86 DD)
Boxwood spider mite – egg hatch (141 DD)
European pine sawfly – larva, early instar (154 DD)
Woolly elm aphid – egg hatch (163 DD)
Inkberry holly leafminer – adult emergence (165 DD)
Spiny witchhazel gall aphid – adult/nymph (171 DD)
Spruce spider mite – egg hatch (179 DD)
Boxwood psyllid – egg hatch (184 DD)
Tea scale – egg hatch / crawler (1st gen) (195 DD)
Hemlock woolly adelgid – egg hatch (1st gen) (197 DD)
Viburnum leaf beetle – first egg hatch (210 DD)
Azalea lace bug – egg hatch (1st gen) (214 DD)
Birch leafminer – adult emergence (219 DD)
Roseslug sawfly – lava/early instar (230 DD)
Elongate hemlock sclae – egg hatch / crawler (1st gen) (232 DD)
Boxwood leafminer – adult emergence (249 DD)

See the Pest Predictive Calendar for more information on DD and plant phenological indicators (PPI) to help you better monitor and manage these pests.

Degree Days (as of April 1, 2026)

Annapolis Naval Academy (KNAK)	113
Baltimore, MD (KBWI)	153
Belcamp (FS836)	118
Clarksville (001MD)	115
College Park (KCGS)	177
Dulles Airport (KIAD)	176
Ft. Belvoir, VA (KDA)	176
Frederick (KFDK)	130
Gaithersburg (KGAI)	155
Greater Cumberland Reg (KCBE)	150
Martinsburg, WV (KMRB)	159
Millersville (MD026)	163
Natl Arboretum/Reagan Natl (KDCA)	197
Perry Hall (C0608)	120
Salisbury/Ocean City (KSBY)	192
St. Mary's City (Patuxent NRB KNHK)	177
Westminster (KDMW)	188

Important Note: We are using the Online Phenology and Degree-Day Models site. Use the following information to calculate GDD for your site: Select your location from the map Model Category: All models Select Degree-day calculatorThresholds in: Fahrenheit °F Lower: 50 Upper: 95 Calculation type: simple average/growing dds Start: Jan 1

Conferences

May 20, 2026
Mid-Atlantic Nursery Workshop: the Water Loop to Pot: Managing Ponds, Irrigation, Substrates, and Runoff for Better Production Profits
Location: Hampton Roads Agricultural Research and Education Center, Virginia Tech, Virginia Beach, VA
For more information on this program

June 16, 2026
2026 Eastern Shore Procrastinators Conference
Location: Zoom

June 18, 2026
MNLGA Field Day
Location: Mt Cuba Center, Hockessin, DE

June 26, 2026
Montgomery County Pesticide Procrastinators Conference
Location: Derwood, MD

IPM Scouts' Diagnostic Sessions (1 - 3 p.m.)
June 23, 2026
July 22, 2026
August 26, 2026
Location: CMREC, Ellicott City, MD

Commercial Ornamental IPM Information
http://extension.umd.edu/ipm

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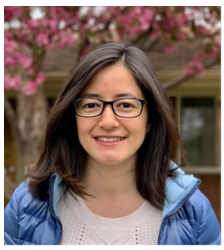

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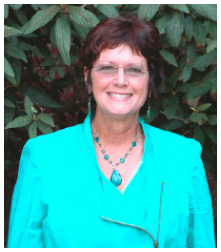

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